

The Significance of Therapeutic Drug Monitoring: Investigating Clinical And Forensic Toxicology

Dr Abanibhusan Jena^{1*}, Dr. Rudra Narayan Pati², Dr. Silky Mahajan³, Dr Seema Yadav⁴

1*Associate Professor and Head of Department, Fakir Mohan Medical College and Hospital, Baleshwer, Odissa Affiliated to: National Medical Commission/FM University Email: drabanibhusanjena@gmail.com 0009-0005-2507-0202

²Assistant professor Department of Pharmacology Fakir Mohan Medical College, Balasore Fakir Mohan University Odisha Email id - rudra.online5200316@gmail.com, 0009-0006-1368-1214

³Designation: Assistant Professor, Department of Pathology College, Punjab Institute of Medical Sciences (PIMS), Jalandhar, Punjab, BFUHS (University) Email id: drsilkymahajan71@gmail.com, 0009-0009-2214-1502

⁴Principal, College of Nursing. Sarojini Naidu Medical College, AGRA, UP. seemakishan22@gmail.com

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Therapeutic Drug Monitoring (TDM) is a vital tool in clinical and forensic toxicology, assessing drug levels in biological samples to optimize therapy or investigate drug-related incidents. TDM, a field that combines clinical pharmacology and forensic toxicology, provides valuable insights into drug therapy management and forensic investigations. This paper studies the role of TDM in clinical conditions, focusing on its effectiveness in guiding dosage adjustments, ensuring therapeutic efficacy, and minimizing adverse effects. The study explores how TDM aids forensic investigations by providing valuable insights into drug-related fatalities, abuse, and compliance monitoring. This study observes current methodologies, challenges, and trends in TDM, emphasizing its crucial role in promoting patient safety, enhancing drug efficacy, and facilitating forensic analyses in toxicology practice. This review studies the role of TDM in clinical conditions, highlighting its potential benefits and limitations in optimizing drug therapy. The widespread adoption and effectiveness of drug metabolism testing are hindered by challenges like variability, assay limitations, and interpretational complexities. TDM is crucial in forensic toxicology for identifying drug-related fatalities, assessing drug abuse patterns, and verifying medication regimen compliance. Postmortem redistribution, analytical sensitivity, and drug concentration interpretation in non-traditional matrices necessitate cautious interpretation and integration with comprehensive forensic investigations. TDM faces challenges in clinical and forensic domains, requiring ongoing research, methodological advancements, and interdisciplinary collaboration to fully realize its potential in patient care and forensic analyses.

Keywords: Therapeutic drug monitoring, Forensic toxicology, Analysis, Pharmacokinetics, Pharmacodynamics

1. Introduction

Human toxicological symptoms have been known since antiquity, with toxins and medications made from plant extracts, animal venoms, and refined mineral combinations. Arsenic, often used in the Renaissance for fortune-seeking, earned the nickname "the inheritance powder." Swiss physician Paracelsus developed the basic theory of toxicology in the 16th century. Forensic toxicology was founded by pioneering work in the 16th and 17th

centuries, with the Marsh test used to determine arsenic poisoning in Charles LaFarge's 1840 murder (Ketha and Garg, 2020). Phases I through IV are developmental steps in creating a novel medication, with Phase II or III studies investigating dosage response and tolerance. However, research on therapeutic drug monitoring (TDM) in Fig. 1. effectiveness for these medications is limited. TDM is useful when drug concentration and effect are strongly correlated, in small therapeutic windows, with no clear clinical parameters, documented

^{*}Corresponding author: Dr Abanibhusan Jena

^{*}Email: drabanibhusanjena@gmail.com

interactions, tracked drug compliance, significant variability in pharmacokinetic parameters (Neef et al., 2008). Hyphenated mass spectrometry (MS) is widely accepted and utilized in the fields of TDM, computed tomography (CT), and Fourier transform (FT). Multiple mass spectrometry (MS) devices can be used in combination with various systems, including chromatography, electrophoresis, MALDI, or paper spray, for varied applications in these domains. Tandem MS, when combined with ultra-high-pressure chromatography (LC), is currently considered the standard method in TDM. HRMS devices are highly versatile and can be used for a wide range of applications, including human toxicity, particularly in CT and FT, as well as TDM. HRMS is expected to gain widespread acceptance due to cost reduction and user-friendly software packages, offering exceptional identification capabilities and simple qualitative and quantitative approach development. Hyphenated mass spectrometry offers superior selectivity and sensitivity, enabling the creation of new techniques and parameter additions. HRMS allows for novel research, reduces analysis time, and discovers new medications, but requires deep understanding and expertise to avoid potential problems (Maurer, 2018). Clinical pharmacology posits that only free medications pharmacologically active, as they interact with specific receptors, and unbound concentrations determine their harmful and effective responses. The equilibrium of a drug's free percentage in plasma and saliva is crucial for assessing medicines in oral fluid, despite over 70 years of studies on saliva's organic solutes. Salivary monitoring requires a consistent, predictable link between saliva and plasma drug concentration, despite various assumptions about saliva drug level monitoring. Measuring oral fluid drug levels can aid in treating patients and adjusting dosages for some medications, but not for most therapeutically supervised medications. Research on antipsychotic drugs suggests that changes in metabolic status, influenced by pharmacogenetic variations or clinical conditions, can be reflected in the parent drug-tometabolite ratio (Langman, 2007). MS is a powerful technology used in research and clinical laboratories for identifying and quantifying compounds. Its identification, high sensitivity, and simultaneous analysis of multiple analytes have led to its rapid expansion in routine clinical practices, particularly in therapeutic drug monitoring, drugs of abuse, and clinical toxicology (Garg and Zhang, 2016, Praveen, 2024). This reviews data on drug stability in blood, plasma, or serum, focusing on newer drugs of abuse and therapeutic drugs. Key information about stability experiments and evaluations is provided. Most drugs are stable under

typical laboratory conditions, except for those with ester moieties or easily oxidized structures. However, specimens should be stored in the refrigerator at -20°C or lower to avoid degradation. Results from biosamples stored at room temperature should be interpreted carefully (Peters, 2007). Over the past 50 years, forensic toxicology has grown dramatically, adding 8-9 divisions. New specimens like hair, oral fluids, blood, and urine can now be used, thanks to the development of instruments like benchtop GC-MS immunoassays. The development of excellent standards and guidelines for medications and poisons in biological specimens has also been a focus of international efforts. Autopsy results, details from the crime scene, and medical history are now taken into account when interpreting toxicological results. An important factor in the advancement of forensic toxicology is societies such as TIAFT (Chung and Choe, 2017, Praveen, 2024). A μ-opioid receptor agonist called methadone is used to treat heroin addiction. Individual differences in its metabolomics have a substantial impact on the toxicological profile and dose response. The liver metabolizes methadone via isoenzymes of cytochrome P450. Comprehending the metabolomics of methadone can aid in the development of customized treatment plans and offer essential case files for legal and medical settings (Dinis-Oliveira, 2016; Buko, 2017). LC combined with MS has become crucial in doping control, clinical and forensic toxicology. Highresolution MS analysis and improved techniques for LC-MS(/MS)-based toxicological studies have improved. Multi-target screening and quantification of medications, toxins, and metabolites are also being explored (Peters, 2011). This essay examines the relationships between clinical and forensic toxicology, focusing on seven areas of analytical toxicology: drug control, brain death, prenatal drug exposure, drug-facilitated crimes, intoxications by new psychoactive substances, and sudden infant death syndrome. Forensic laboratories investigate situations like SCD, SIDS, and doping control, while clinical laboratories handle issues like FAS and drug exposure during pregnancy. Both fields share common topics, fostering communication and enhancing the expansion, dependability, and robustness of both types of laboratories (Barcelo et al., 2018). The findings of a comprehensive study that investigated the varied applications of TDM in forensic and clinical contexts are presented in this article. The outcomes highlight the critical role that TDM plays in modern forensic research and healthcare, providing insightful information on toxicological analyses, drug therapy management, and medico-legal questions. Clinicians and forensic scientists can expand the study of drug-related

phenomena in both clinical and legal contexts, improve drug safety, and improve patient care by utilizing TDM methodology and technology.

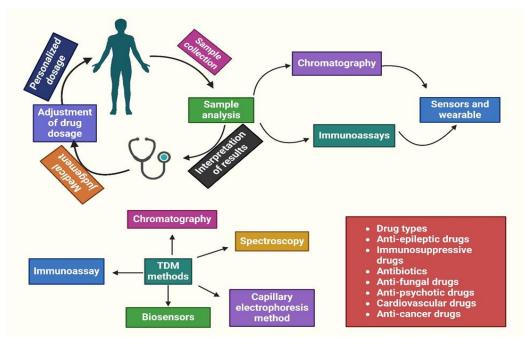


Fig. 1. Therapeutic Drug Monitoring Process and Methods

The rapid growth of clinical pharmacy in China has led to the importance of therapeutic drug monitoring (TDM), a strategy that optimizes individualized drug therapy by combining pharmacokinetic and pharmacodynamic knowledge. TDM aims to reduce drug-drug toxicity, prevent drug resistance, and improve treatment outcomes. Accurate analytical procedures are crucial for successful implementation.

2. Pharmacokinetics and pharmacogenetics2.1. Pharmacokinetic aspects

2.1.1. Absorption, distribution, and elimination of neuropsychiatric drugs

This list of typical pharmacokinetic characteristics does not include all of the outliers. Aripiprazole and fluoxetine, on the other hand, have lengthy elimination half-lives (3-15 days for fluoxetine and 72 hours for aripiprazole, accounting for its active metabolite norfluoxetine). Examples of medications with short elimination half-lives tranylcypromine, trazodone, venlafaxine, agomelatine. Patients with weak liver function may benefit from the fact that sulpiride, gabapentin, memantine, milnacipran, or amisulpride primarily eliminated really and only poorly metabolized in the liver. The reason for the nonlinear pharmacokinetics of paroxetine is that it inhibits its metabolism by the binding of a metabolite that inactivates the enzyme irreversibly (Bertelsen et al., 2003). The enantiomers of many neuropsychopharmacological medications, which are utilized as racemic substances, have markedly different pharmacodynamic and pharmacokinetic characteristics (Baumann et al., 2002, Smith, 2009). However, TDM of the enantiomers has only been introduced for two racemic psychoactive substances thus far: methadone and methylphenidate (Balant et al., 1989, Eap et al., 2002). Racemic methylphenidate has an active enantiomer called (R)-methadone. Its therapeutic action is mainly attributed to l-methylphenidate, also known as levorotary methylphenidate. Flupentixol decanoate in its depot formulation contains only the cis-isomer of flupentixol. In contrast, the oral form of flupentixol is administered as an equal mixture of the geometric isomers, specifically the cis- (Ztrans- (E-isomer) forms. As isomer) and demonstrated by clinical investigations, the latter is the only one deemed pharmacologically active in terms of its affinity for receptors of serotonin and dopamine. Cis-flupentixol efficacy (Z-flupentixol;αflupentixol) appears to be greater than the effect of trans-flupentixol (Baumann et al.,, 2012). The different activities of enzymes involved in the metabolism of drugs are the cause of inter- and intra-individual variations in concentrations of neuropsychopharmacological medications in the blood. Age may cause a decline in enzyme activity (Klotz, 2009), and hepatic and renal disorders may alter it. CYP enzymes are primarily responsible for catalyzing phase 1 reactions. These proteins are members of a superfamily that act as terminal oxidases in electron transfer chains and include

heme as a cofactor. The spectrophotometric peak at 450 nm, which is the peak wavelength at which CYP enzymes, when in their reduced state and bound to carbon monoxide, absorb the lightest, is the source of the word P450. Phase 1 reactions, catalyzed by CYP enzymes, introduce polar functional groups, allowing for subsequent phase 2 reactions where highly polar substances like sulfuric acid or glucuronic acid are conjugated. Glucuronidation of a hydroxyl group (as seen with oxazepam or lorazepam) or an amine group to form Nglucuronides (such as with olanzapine) is a key metabolic pathway neuropsychopharmacological drugs with these functional groups. The CYP enzyme family is organized into 18 families and 43 subfamilies based on their amino acid sequences. In humans, different gene clusters encode 57 functional CYP genes and 58 pseudogenes (Zanger and Schwab, 2013). Important isoenzymes for the metabolism of neuropsychopharmacological drugs are CYP1A2, CYP2B6, CYP2C8, CYP2C9, CYP2C19, CYP2D6, CYP2E1, and CYP3A4/5 (Zanger and Schwab, 2013; Backman et al., 2016; Zhou, 2009; Zhou et al., 2009). It is possible that additional enzymes as metabolic key factors function pharmacological and harmful effects of drugs (Barski,). Ketone or aldehyde group reduction in endogenous and exogenous substances is catalyzed by enzymes belonging to the AKR superfamily, including aldo-keto reductases (AKRs). Thirteen AKR proteins have been found in humans (Barski et al., 2008). It was demonstrated that they convert naltrexone to naltrexol (Breyer-Pfaff and Nill, 2004) and ziprasidone to its dihydro derivative (Beedham et al., 2003). Citalopram undergoes stereoselective deamination by monoamine oxidase subtypes A and B (MAO-A and MAO-B), resulting in an acidic metabolite that appears to be inactive (Rochat et al., 1998). In actuality, phase 2 enzymes are becoming more and more characterized in terms of substrate selectivity. Regarding their affinity for substrates, the isoenzymes overlap significantly (Court, 2010; Oda et al., 2015). The liver is the primary site of drug metabolism, with some metabolism also occurring in extrahepatic tissues. like the brain or intestinal mucosa (Benedetti et al., 2009; Gervasini et al., 2004; Meyer et al., 2007). Pharmacokinetic drug-drug interactions may arise when medications that inhibit or induce drug-metabolizing enzymes are combined (Abernethy et al., 1985), provided that the medication is a substrate of the enzyme that is either inhibited or stimulated. TDM has discovered numerous interactions through either accidental or retrospective examination of TDM databases (Castberg et al., 2007; Hefner et al., 2015; Paulzen et al., 2016; Rasmussen and Brøsen, 2000). Smoking is one of the environmental factors that has the

greatest therapeutic significance for medications that are CYP1A2 substrates (Ereshefsky et al.,, 1985; Faber et al.,, 2005) Polycyclic aromatic hydrocarbons of cigarette smoke stimulate CYP1A2 in a dose-dependent manner. CYP1A2 activity is enhanced by 1.2, 1.5, and 1.7 while 1-2, 6-10, and more cigarettes smoke each day, respectively (Faber and Fuhr, 2004). Three days after quitting smoking, the elevated activity returns to baseline. Therefore, when smoking more than ten cigarettes a day, smoking consequences should be taken into account (Faber et al., 2005). When using a CYP1A2 substrate in Table 1, for therapy, such as olanzapine (Zullino et al., 2002), duloxetine (Fric et al., 2008), or clozapine (Bondolfi et al., 2005) (van der Weide et al., 2003), stopping heavy smoking may need dose decrease, which TDM should regulate. Drug transporters are engaged in the distribution of pharmacokinetics of pharmaceuticals in addition to Enzymes participating in metabolism phase 1 and phase 2 (Bruhn and Cascorbi, 2014; Dong et al.,, 2009; Ufer et al., 2011; Wolking et al., 2015). These proteins, called ATP-binding cassettes (ABCs), are found in membranes of cell, and serve as efflux transporters that shield organs from outside substances. The major factors that determine the distribution kinetics of neuropsychopharmacological drugs have been identified as ABC transporters including breast cancer resistance protein (BCRP) which is encoded by ABCG2, multidrug resistance protein (MRP) which is encoded by ABCC1 and P-glycoprotein (Pgp) which is the gene product of ABCB1 (Wolking et al., 2015). Substances that are ABC transporters substrates enter cells by passive diffusion and are then released into the extracellular space by ABC transporters through conformational changes that are dependent on ATP. Due to its strong expression in the small intestine and the blood brain barrier (BBB), P-gp is (Wolking et al., 2015) a crucial regulator of drug trafficking into and out of several organs (Wolking et al., 2015). Research on animals indicates that P-gp regulates the brain's availability rate of numerous antipsychotic and antidepressant medications, such as risperidone, citalopram, and nortriptyline (Doran et al.,, 2005; Suzuki et al.,, 2014; Uhr et al.,, 2003). It has been proposed that ineffective concentrations are caused by high P-gp function, and tolerability issues and high drug concentrations are linked to poor P-gp activity (Bet et al.,, 2016; Breitenstein et al.,, 2015; Breitenstein et al., 2016; Brückl and Uhr, 2016; De Klerk et al., 2013, Nikisch et al., 2011; Ray et al., 2015; Praveen et al., 2024; Uhr et al., 2008). ABC transporters have been found to have several genetic mutations, much like CYP enzymes (Wolking et a., 2015). Furthermore, there are numerous mechanisms to up- or down-regulate the expression of ABC

transporters, including pathophysiological stresses, xenobiotics, hormones, and nutritional factors 2015). The pharmacokinetics neuropsycho pharmacological medications have also been found to differ between genders (Aichhorn et al., 2006; Marazziti et al., 2013, Sigurdsson et al., 2015; Soldin and Mattison, 2009). This is most likely because female sex hormones have an impact on the absorption, distribution, metabolism, and excretion of pharmacokinetic processes (Damoiseaux et al., 2014; Kokras et al., 2011). The results continue to be inconsistent, and it's unclear how applicable they are clinically. Some investigations indicated that the influence of body mass was lower than expected by pharmacokinetic principles (Aichhorn et al.,, 2006, Sigurdsson et al.,, 2015, Soldin and Mattison, 2009, Praveen and Morales-Bayuelo, 2023), despite the fact that body mass should be a key predictor of the concentration of a medicine in blood following administration of a given dose (Steimer, 2004). In these domains, systematic research is still necessary.

2.1.2. Blood drug concentrations

Drug intake equals drug elimination during a predetermined period of time at a steady state. Concentrations will change during the day, particularly for medications having brief elimination half-lives (<12 hours) and based on the dosage regimen, which is important to take into account when interpreting TDM data (Steimer, 2004). For the great majority of medications in TDM, trough steady-state (treatment with continuous dose for not less than 4 to 6 half-life) concentrations (Cmin) have been the accepted practice. For practical reasons, the method of trough sampling just before the subsequent dose has been selected. The time curve of concentration is almost horizontal towards the end of the dosing period (terminal β-elimination phase) and deviations from the right sampling time immediately before the next dose are less critical for trough samples than for the other phases after dose administration.

2.2. Pharmacogenetic features

More and more research has shown the clinical significance of pharmacogenetic variables in the pharmacodynamics and pharmacokinetics of neuropsychiatric medications (de Leon, 2006; Evans and M. V. Relling, 1999; Mrazek, 2010; Samer et al., 2013). Drug-metabolizing enzymes, particularly CYP isoenzymes, display genetic diversity, as was previously reported (Zhou, 2009; Zhou et al., 2009). Wild-type individuals with two active alleles are referred to as extensive metabolizers (EM). PMs are those who do not have functional alleles. Genetically heterozygous for both inactive and active alleles, or with one or two alleles

with decreased activity, are the two types of intermediate metabolizers (IM). Alleles with enhanced activity or functional allele multiplications are carried by ultrarapid metabolizers (UM) (Bergmann et al., 2001). Clinical significance arises from drug-metabolizing enzyme genetic variations. On the one hand, elevated blood concentrations in PM can lead to unforeseen toxicities and severe medication reactions. Conversely, subtherapeutic blood concentrations in UM may lead to nonresponse (Jose de Leon et al., 2005). CYP enzymes participate in the metabolism of prodrugs, such as CYP2D6, which converts morphine from codeine and desmethyltramadol from tramadol (Ortiz de Montellano, 2013; Huttunen et al., 2011). Under these circumstances, PM patients won't have the capability to generate metabolites that pharmacologically active, and UMs run the danger of experiencing unpleasant medication reactions. Determining the mRNA encoding CYP1A2, CYP2C9, and CYP2C19 in leukocytes is a novel and promising method. Parallel probe drug phenotyping of CYP enzymes demonstrated a strong correlation between mRNA levels and hepatic CYP activity (Temesvári et al., 2012). In the past, probe medications like midazolam for CYP3A4/5, metoprolol or dextromethorphan for CYP2D6, omeprazole for CYP2C19, or caffeine for CYP1A2were used to ascertain the metabolizer status. (Tanaka et al., 2003; Liu et al., 2009; Skogh et al., 1999). These phenotyping assays allow for the detection of metabolic alterations by measuring the patient's metabolic state during the test. Thus, they can be used to investigate how CYP activities are affected by environmental factors like smoking or prescription drugs (Faber et al., 2005; Zullino et al., 2002; Skogh et al., 1999). The availability of CYP genotyping has increased during the past few years. The undeniable benefit of genotyping is that it serves as a "trait marker" and is unaffected by external circumstances. Its outcome has a lifelong worth and can be executed in any circumstance. However, the presence of rare genetic variations contributes to a notable amount of variation, making it possible to forecast one's enzyme activity by genetic analysis that specifically focuses on prevalent alleles (Matthaei et al., 2015). This is true even though the functioning importance of the genetic variants for CYP enzymes has been thoroughly described (Gaedigk et al., 2008). Though their clinical significance in pharmacotherapy and for adjusting doses is not as clearly established as that of CYP polymorphisms (Stingl et al., 2003), alternative metabolic systems of enzymes, like UDP glucuronosyltransferases (UGT), also exhibit genetic variations (Court 2010, De Leon, 2003). It has been proposed that the ABCB1 genotype influences the response to antidepressant and

antipsychotic drugs due to its involvement in ABCB1 transporters and the gene product P-gp has an essential function in the transport of drugs throughout the body. Antidepressant treatment outcomes can be enhanced by ABCB1 genotyping, and individuals may exhibit varying responses to antidepressants that are substrates of P-gp. In the meanwhile, more than 30 studies have looked into the possibility that human antidepressant clinical efficacy and/or tolerability may be predicted by genetic variations in ABCB1. Antidepressant effects have been reported to affect minor allele carriers more frequently than major allele carriers of specific polymorphisms single nucleotide (SNPs): rs2032583 and rs2235040 (Breitenstein et al.,, 2015; Breitenstein et al.,, 2016; De Klerk et al.,, 2013; Ray et al., 2015; Uhr et al., 2008; Roberts et al., 2002; Sarginson et al., 2010). However, several additional studies (Dong et a., 2009; Bet etal., 2016, Perlis et al., 2010; Schatzberg et al., 2015) did not find that minor allele carriers had more adverse medication reactions or greater response rates than noncarriers. Carriers of the minor allele of rs2235083 had greater efficacy at doses within the optimal dosage range in an initial clinical trial using various of P-gp substrates antidepressants (Breitenstein et al., 2016; Brückl and Uhr, 2016). Apart from the pharmacokinetic elements discussed earlier, there is mounting evidence that genetic variables influencing pharmacodynamic processes—like how drugs interact with enzymes, transporters, receptors, structural proteins, or ion channels—are essential in determining how well a treatment works for mental health conditions. The most studied gene in relation to affective disorders is the serotonin transporter gene (5HTT; SLC6A4). But the results are still inconclusive (Kato et al., 2015; Serretti et al., 2007; Taylor et al., 2010). GWAS has been conducted using a hypothesis-free approach on the STARD, MARS, and GENDEP datasets. However, these investigations were unable to find genome-wide significant markers of response to antidepressant treatment (Hohmann et al., 2015, Laje and McMahon, 2011). The greatest meta-analysis to date has examined the response to lithium in a cohort of over 2,500 individuals from 22 research centers worldwide. The findings are not currently significant for clinical decision making, even though they could serve as a foundation for a deeper comprehension of lithium mechanisms (Hou et al., 2016; Laje, 2013; McCarthy et al., 2010; Schulze et al.,, 2010). The DRD2, DRD3, and DRD4 genes have been widely studied in relation to psychotic disorders and their response antipsychotic treatments. But these studies have not been able to produce consistent replicable results (for a review see Brandl et al., 2014). Recent metaanalysis studies suggest that the A118G

polymorphism of the µ opioid receptor gene (OPRM1) is a significant predictor of the response to naltrexone in alcohol-dependent patients (Chamorro et al., 2012). Future studies are needed to establish the clinical effectiveness (e. g., diagnostic accuracy, positive and negative predictive values) of pharmacogenetic testing for alcohol use disorders based on OPRM1 genotypes (Hendershot, 2014). At the pharmacodynamic level, pharmacogenetic investigations produced encouraging preliminary findings about the underlying genetics of pertinent adverse drug reactions to psychoactive medications. Patients of Asian descent who are treated with carbamazepine have a consistently greater chance of developing Stevens-Johnson syndrome when their human leukocyte antigen markers, HLA-B1502 and HLA-A3101, are present (Wu et al., 2015; Ferrell and McLeod, 2008). A few pharmacogenetic assays, such the PGxPredict: CLOZAPINE test, which was intended to predict the risk of agranulocytosis based on HLA-DQB1 gene variant, were tested in clinical settings. Despite having a high specificity of 98.4% and a poor sensitivity of 21.5%, the test has since been discontinued (Ho and Reddy, 2011). Antipsychotic-induced weight gain has been demonstrated to be mediated by 5-HTR2C, leptin gene variants, neuropeptide Y (NPY), melanocortin 4 receptor (MC4R), cannabinoid receptor 1 (CNR1), and neuropeptide Y (NPY) (for review see Gressier et al., 2016). In antipsychotic-induced dystonia/tardive dyskinesia, well-replicated gene variations have been reported: variations in the serotonin receptor genes HTR2C (Al-Janabi et al., 2009; Hadithy et al., 2009; Segman et al., 2000) and possibly also HTR2A (Lerer et al., 2005; Segman et al., 2001), as well as variations in RGS2 (regulator of G-protein signaling 2), a gene that modulates dopamine receptor signal transduction (Greenbaum et al., 2009; Greenbaum et al., 2007) . Negative symptoms in schizophrenia that respond well to antipsychotic treatment are linked polymorphism in the serotonin receptor gene HTR1A (rs6295; C-1019G) (Mössner et al., 2009; Takekita et al., 2016). In an effort to address the shortcomings of earlier research, the following tactics have been suggested: Concentrating on a single pharmacologic class and precisely delineated phenotypes (e.g., ISPC (Biernacka et al., 2015, Azad et al., 2024), factors related to the environment (Klengel and Binder, 2013) and pharmacokinetic parameters (e.g., blood levels (Proft et al., 2014; Unterecker et al., 2015), Enhancing genetic analysis by incorporating structural variation (e.g., copy number variation (O'Dushlaine et al., 2014), examining the combined effects of multiple risk genes ('epistasis', e.g., (Mas et al., 2015; Domschke et al., 2014), and incorporating epigenetic variation

(Domschke et al., 2014; Menke et al., 2012). In this vein, sizable global consortia, such as the International Consortium on Lithium Genetics (ConLiGen) (Schulze et al., 2010), are being formed in an effort to carry out extensive pharmacogenetic research using cutting-edge methods like exome sequencing and genome-wide association studies.

3. Utilizing blood drug concentrations for Neuropsychopharmacotherapy direction

TDM considers both pharmacodynamic and pharmacokinetic factors to direct neuro psychopharmacotherapy. To ensure therapeutic efficacy and acceptable tolerability, it is necessary to determine whether the concentration of a drug is within the therapeutic reference range and, if so, whether the blood concentration matches the recommended dosage. The latter determines whether the medication is taken as directed and, if not, whether there are irregularities in the pharmacokinetics. As such, it is necessary to distinguish between predicted dose-related drug concentrations and therapeutically effective drug concentrations (Haen, 2011, Haen et al., 2008).

3.1. The range of therapeutic reference

Pharmacologic effects are thought concentration-related according to the rule of mass action (Aronson and Ferner, 2016). This premise underpins TDM in terms of both therapeutic betterment and unfavorable drug responses. TDM also presupposes that an array of medication concentrations in blood, known as the "therapeutic reference range," are necessary for both acceptable safety and maximum efficacy. Since the 1960s, research on the connections between blood drug concentration and clinical improvement has substantiated this idea about antidepressants that first-generation are tricyclic, lithium, and antipsychotic medications. Nortriptyline, imipramine, and desipramine—drugs linked to a high probability of response—have been shown to have a substantial correlation with blood drug concentration in connection to clinical outcomes in meta-analyses and systematic reviews based on welldesigned research (Baumann et al., 2004). A metaanalysis of 45 studies using amitriptyline as a model different molecule revealed that statistical techniques produced nearly equivalent therapeutic reference ranges (Ulrich and J. Läuter, 2002; Ulrich et al., 1998). New antipsychotic medications such as olanzapine (Perry et al., 1997), risperidone (Yasui-Furukori et al., 2010), and aripiprazole (Sparshatt et al., 2010) have been shown to have connections between their blood concentration and clinical effectiveness (Lopez and Kane, 2013). When using TDM-guided medication, the therapeutic reference range is a critical zone that needs to be targeted.

Determining the lower and upper bounds of medication concentrations in the blood that are both therapeutically efficacious and tolerated is necessary for its estimate. There isn't a widely used technique to calculate these limits, methodological constraints like treatment resistance or placebo response need to be taken into account (Aronson and Ferner, 2016; Preskorn, 2014). The terms "orienting therapeutic range," "target concentration," "target range," "effective plasma concentration," "optimal plasma concentration," "therapeutic range," and "therapeutic window," which was the phrase utilized in the initial TDM consensus (Baumann et al., 2004), are many synonyms for "therapeutic reference range." The AGNP TDM task force determined in 2011 to employ the phrase "drug concentration in blood," which encompasses plasma level or serum concentration, plasma concentration, blood level or serum level, and to use the term "therapeutic reference range," adhering to the publication of TDM recommendations for antiepileptic medicines is a common practice (Patsalos et al., 2008). The evidence-based therapeutic reference ranges were obtained from the literature through the abovedescribed structured review approach. Only 17 neuropsychiatric medications had therapeutic reference ranges according to randomly assigned clinical studies identified in the literature. Reference ranges for the majority of medications came from research using dosages that were therapeutically efficacious. Generally speaking, the reference ranges for the principal indication are those found in. However, numerous medications are advised for multiple purposes. For instance, antipsychotic medications are licensed for the treatment of affective disorders, and antidepressant medications are used to treat anxiety, obsessive compulsive disorder, and chronic pain. The ideal blood medication concentrations for various indications Carbamazepine, well understood. lamotrigine, and valproic acid (valproate) are the exceptions, and as a result, they are occasionally included twice. Research to assess therapeutic reference ranges for patients who are juveniles or adolescents is currently underway (Egberts et al., 2011; Gerlach et al., 2016; Koelch et al., 2012; Taurines et al., 2013; Wohkittel et al., 2016).

3.1.1. Determining the therapeutic reference range's lowest limit

When feasible, research assessing the correlation among a drug's blood concentration and clinical efficacy should serve as the foundation for determining the lower bound of the therapeutic range. The effects of the medication are not appreciably different from a placebo below the minimum threshold. A potential study that follows

a double-blind trial with a randomized control with patient doses that produce a predetermined range of blood concentrations of the drug is the best study design to assess the minimum threshold. It used an essentially perfect research design on clozapinetreated individuals (VanderZwaag et al., 1996). The blood was titrated to 50-150 ng/mL, 200-300 ng/mL, or350-450 ng/mL clozapine concentrations. When compared to low quantities of clozapine, intermediate and high concentrations showed a significant therapeutic advantage. Blood level research contrasting mirtazapine imipramine was conducted using a similar approach (Bruijn et al., 1996). However, carrying out these investigations presents a significant logistical difficulty. For the assessment of the lower limit, fixed dosage studies are more practical and desirable (Ulrich and Läuter, 2002; Ulrich et al., 1998). Receiver operating characteristic (ROC) analysis has shown usefulness in estimating the therapeutic reference range lowest value (Hanley and McNeil, 1982). A ROC plot analyzes the accuracy and precision of the variable "drug concentration in blood" and enables the determination of a cut-off value that distinguishes responders from nonresponders. For some antipsychotic antidepressant medications, the value of ROC analysis has been established (Müller et al., 2007; Perry, 2001, Perry et al., 1994; Waldschmitt et al., 2009).

3.1.2. Determining the therapeutic reference range's top limit

An inverse U-shaped connection among clinical activity and blood concentrations was observed for nortriptyline in the first investigation on TDM in psychiatry (Åsberg et al., 1971). The tricyclic antidepressant drug's method of action on monoaminergic neurons was blamed for the lack of therapeutic improvement at high dosages. But based on what is now known, it is more plausible that the side effects of nortriptyline are what are causing the decreased amelioration at high dosages. As a result, even under these guidelines, the upper bound of the therapeutic range is frequently determined by the elevated risk of adverse medication reactions. Antipsychotic medication motor symptoms (Rao et al., 1980) and tricyclic antidepressant drug side effects (Dawling, 1982; Gupta et al., 1999) have been linked to blood drug concentrations. The blood content of paroxetine was observed to positively correlate with symptoms of serotonin syndrome (Hegerl et al., 1998). It was demonstrated that the clearance of citalopram was inversely linked with adverse medication responses (Yin et al., 2006). ROC analysis can be used to determine the top limit of the therapeutic range when such data are available (Müller et al., 2007). However, there is

insufficient reliable data regarding the blood levels and the frequency of adverse drug reactions for several of the neuropsychiatric medications included. Most case reports on intoxications or tolerability issues omit measurements of drug concentrations. Reports of intoxications and fatal instances that appear sporadically are not very useful. The medication level is typically significantly above the threshold linked to optimal therapeutic effects when recorded blood concentrations have resulted in death (Reis et al., 2007; Stead and Moffat, 1983). Additionally, post mortem drug redistribution into or out of the circulation can cause abrupt changes in blood levels (Kugelberg et al., 2004; Pounder and Jones, 1990), and the change's direction is not always consistent (Kennedy, 2010).

3.2. The reference range linked to dosage

A second concentration range, known as the dose-related reference range, exists in addition to the therapeutic reference range for the purpose of interpreting TDM results. The therapeutic reference range is a pharmacodynamic method that is used. Pharmacokinetics is the application of the dose-related reference range. It makes a comparison between a drug's measured concentration and its theoretically predicted range. These studies are preferably conducted on a population of normal patients. In pharmacokinetic studies, the average steady-state concentration (Cav) of a drug in a normal patient can be determined when we know the bioavailability (F), total clearance (CL), dosing interval (di), and daily maintenance dose (Dm),

$$Cav = (F/CL) \times (DM/di) - ... (1)$$

The prescription specifies the dosage and the interval between doses, while pharmacokinetic trial data provides the pharmacokinetic parameters. Cav±SD (ng/mL) can be calculated by Eq. (1) utilizing the standard deviation (SD) of the total apparent clearance CL/F (mL/min), the daily dose (1 mg/24 h = 106 ng/1440 min). The dimensions of the various factors must be taken into account throughout the computation, and all doses, volumes, and time periods must be translated to ng, mL, and min, respectively. The coefficient of variation is 50% when the CL/F value is provided as 100±50 mL/min. This means that for a dose of 20 mg/day,

$$Cav = \left(\frac{20,000,000 \, ng}{1440 \, min}\right) \left(\frac{1ml}{100 min}\right) = 139 \, \text{ng/mL}$$

The standard deviation is 69 ng/mL. So, Cav±SD varies from 70 to 208 ng/mL. By Haen and colleagues (, Landmark et al., 2016), the Cav±SD range was recommended as a dose-dependent

reference range, assuming a 24-hour dosing interval. Within this range, the mean - SD was regarded as the lowest bound and the mean + SD as the maximum threshold. According to statistics, this range includes 68% of the quantities seen in the blood of a population of people aged 18 to 65 under typical circumstances. Apparent total clearance data±SD for 83 neuropsychiatric medications were taken from the literature for the 2011 guidelines (Hiemke et al., 2011) in order to calculate dosage factors. Dose-dependent reference ranges were computed and utilized for the analysis of TDM findings by multiplying the daily dose with these factors ± SD. A patient's drug concentration was considered normal when it was determined by TDM to be within the reference range associated to their dosage. Concentrations that were either above or below the range were seen as indicators of possible anomalies, such as partial non-adherence, interactions between drugs, genetic variations in the enzymes that metabolize drugs, or illnesses of the organs that remove drugs.

The dose-dependent reference range concept was effective. It was possible to identify a large number of individuals who were either pharmacokinetically abnormal or incompletely adherent (Haen, 2011). When the drug's elimination half-life (t1/2) is longer than the dosage frequency, the equation for Cav is accurate and helpful. Nevertheless, values determined by Eq. (1) are not very indicative of the Cmin values used for TDM when the half-life is brief and the time between doses is larger than the half-life. With a t1/2 of 14 hours and valproic acid administered once or twice daily.

The dose-related reference range of valproic acid, as determined by Eq. (1), is 94±35 µg/mL when administered daily in dosages of 900 mg, regardless of the interval between doses. Time to concentration curves, on the other hand, indicate that if the dosage regimen consists of a single 900 mg dose every day, the concentrations in the trough are less than Cav, 49±15 μg/mL. If a regular dosage of 900 mg per day is given in two equal doses of 450 mg each, it equals 69±25 µg/mL. For dosage intervals less than 14 hours, the Cav±SD and Cmin±SD ranges correspond. As a result, calculated Cav can be regarded as a reliable indicator of the anticipated concentration of drugs in the bloodstream. However, Cmin is 54% lower than Cav 24 hours after the last dosage while following a single dose per day schedule. This constraint needs to be taken into account when utilizing Eq. (1) based computations of dose-related reference ranges, as valproic acid is used as an example to illustrate. This restriction may apply to several medications, such as naltrexone, atomoxetine, venlafaxine, paroxetine, or duloxetine, depending on the dose interval. The values calculated are reduced by over 30% for Cmin than for Cav when dosage intervals exceed t1/2. In total, 32% of the compounds included in fall into this category.

Additionally, there exists an additional constraint for Cav-based computations. TDM is predicated on measuring the minimum concentration of a drug in bloodstream; in contrast, Cmin allows for easy measurement verification of the accuracy of the dose-related reference range. Cav is defined as the area under the dose interval divided by the area under the time to concentration curve (AUC). It cannot be linked to a specific moment in time, such as Cmin, which is required for venipuncture timing. Neglecting daily variations in drug levels, which might be significant for a medication's tolerance and efficacy (Chenu et al., 2009), is another drawback of Cav-based computations.

It was determined to alter the computation of dose-related reference ranges for this update due to these constraints. By extending Eq. (1) and using the Bateman function, steady-state concentrations can be computed without delving into the specifics covered in pharmacokinetics courses (see, for example, Bauer et al., 2008; Dost, 1953). Using this method, Gex-Fabry and colleagues (Gex-Fabry et al., 2003) calculated concentration throughout the elimination phase and provides an objective for the postabsorptive phase, which refers to the period among tmax, the time when the concentration of drug is at its highest, and tmin, Cmin.

For every time point during the postabsorptive phase, a predicted steady-state drug concentration Ct can be calculated as follows, considering a model with only one compartment and a decline that follows an exponential pattern of the concentration of drugs in the bloodstream:

$$Ct=(e^{-ke\times t}) \times [(ke\times di)/(1-e^{-ke\times di})] \times [(F/CL) \times (Dm/di)]$$
 -----(2)

where t is blood withdrawal time, $t_{1/2}$ is elimination half-life, ke is elimination rate constant, di is dosing interval, CL/F apparent total clearance, and Dm is the dose under steady state conditions

$$ke = \frac{ln2}{t_{1/2}}$$

Using Eq. (3), one may estimate an expected Cmin as follows, assuming, di = 24 h, t = time,

 Δt = Duration between the most recent consumption of medication and the withdrawal of blood.

Cmin=
$$(e^{-ke \times \Delta t}) \times [(ke \times 24)/(1-e^{-ke \times 24})] \times (F/CL) \times (Dm/24)$$
 -----(3)

It is therefore possible to calculate the drug concentrations predicted by TDM measurements using the values.

For medications with known CL/F and $t_{1/2}$, a DRC factor can be constructed & calculated, for example, using MS-Excel software, by using part of Eq. (3).

DRC factor =
$$(e^{-ke \times \Delta t}) \times [(ke \times 24)/(1-e^{-ke \times 24})] \times (F/CL)$$
 (4)

Then, by calculating the product of the DRC factor and the daily dose, one can determine the expected Cmin of a given dose. The necessity to apply $t_{1/2}$, that as well differs throughout persons, and the more involved calculation process are the limits for predicting theoretically predicted Cmin in contrast to Cav. The TDM guidelines regard the standard deviation of mean drug concentrations in patients who comply as the normal variability of perceived total clearance, which is probably attributable to comparable reasons. This presumption led to the definition of the relationship between the variation between individual's CL/F and the variation of the Cmin. As was previously done for Cav-based estimates (Haen et al., 2008), the SD provided in the literature for CL/F was thus propagated to Cmin to determine anticipated mean±SD as dose-related reference range. The prediction of expected drug concentrations using this method of calculation was evaluated empirically.

The DRC variables for 172 compounds, including parent medicines, metabolites, and active moiety. Eq. (4) was utilized to calculate factors by utilizing pharmacokinetic data that was published in articles. Recommendations for drug administration regimens were followed for defining Δt . It was 24 hours for a medication like citalopram or extended release (XR) venlafaxine administered once daily in the morning. It was set at 12 hours for medications such as amitriptyline, which is generally administered in the morning and evening. It was fixed at 10 hours for hypnotic medications administered just before bed and blood withdrawal the following morning. By multiplying the DRC factors low (= DRC factor – SD) and high (= DRC factor + SD) by the daily dose, the above factors can be used to calculate the lower and upper limits of the range and to produce the dose-related reference range. DRC factors for Δt at 12 and 24 hours, respectively, for medicines administered once or

twice daily. Since blood concentrations are not determined at tmin (no trough levels) for medications such as modafinil or clomethiazole, DRC factors are provided at specific times when blood withdrawal is advised.

For each time point in the postabsorptive phase, anticipated concentrations of drugs can be calculated using Eq. (2) when Δt deviates from the values given.

The dose-related reference range, which was first introduced as the dose-related reference range for average drug concentrations (Haen et al., 2008), is currently understood to be a Cmin range that may be computed using the recommended dosage and pharmacokinetic characteristics.

3.3. Ratio of concentration to dosage

An additional metric to analyze pharmacokinetic anomalies is the proportion of medication concentration to dosage (Cmin/D, sometimes shortened as C/D) (Leon et al., 2013, Hefner et al., 2013; Diaz et al., 2008). C/D can be computed with ease using TDM data. Total clearance has an inverse relationship with C/D ratios ((Leon et al., 2013, Hefner et al., 2013; Diaz et al., 2008)). Drug clearance is accelerated by a low C/D ratio and sluggish by a high C/D ratio. By comparing several patient groups, C/D ratios were utilized to identify drug-drug interactions (e.g., (Paulzen et al., 2016, Burns et al., 2016; Schoretsanitis et al., 2016). Jerling and colleagues assessed intraindividual C/D ratios of nortriptyline and amitriptyline and discovered interaction activity of carbamazepine, perphenazine, levomepromazine and demonstrating that earlier C/D values substantiated both on and off concurrent medications (Jerling et al., 1994). As was demonstrated with clozapine (Stieffenhofer et al., 2011), repeated monitoring in the exact patients aids in the detection of partial medication nonadherence. The C/D intraindividual variability should be less than 20%. If the variation is more than 20%, it may indicate pharmacokinetic changes or issues with adherence brought on by interactions between drugs, foods, and diseases. The C/D ratio can also be used to calculate the dose needed to get the drug's blood concentration down to the desired target (Armijo, et al., 1997).

4. Clinical applications

In the clinical laboratory, immunoassays are frequently used techniques for tracking both medicinal medications and drugs of abuse. Due to the possibility of false positive or false negative results from immunoassays. Because immunoassays may not be available for certain medicines and lack

specificity or cross-reactivity, mass spectrometry has been used to confirm immunoassay results (Maurer et al., 2007) and occasionally as a screening tool. Small molecule drug measurement is one of the primary factors propelling mass spectrometry's growing number of clinical practice applications and is also pushing the boundaries of technology development. This volume focuses on toxicology and therapeutic medication monitoring. Another early adopter of mass spectrometry was testing for the confirmation and screening of inborn errors of metabolism, which has been crucial in expanding the uses of mass spectrometry (Garg and Dasouki et al., 2006, Jones and Bennett, 2002). Many advancements have recently been made in novel research areas, especially in the fields of endocrinology and hormone testing in clinical laboratories (Pagotto et al., 2013; Vogeser and Parhofer, 2007; Soldin and Soldin, 2009). Applications of MS are growing in the analysis of big molecules, including peptides, proteins, lipids, polysaccharides, and DNA, even if these are not yet often seen in clinical laboratories (Jimenez and Verheul, 2014; Li et al., 2014; Whiteaker, 2010). The use of matrix-assisted laser desorption/ionization (MALDI) mass spectrometry for quick bacterial identification is another developing field (Ho and Reddy, 2011; Lagacé-Wiens, 2015; Luan et al., 2009).

5. Clinical and forensic toxicology screening

If the proportion of positive outcomes is minimal and large amounts of samples need screening in Fig. 2, IA prescreening for a restricted number of substances of abuse designed for workplace drug testing in the USA is still widespread in CT and FT. Because most NPS are not detectable by IA, and because positive results need to be validated in cases of high prevalence (Yuan et al., 2015; Petrocheilou et al., 2017), hyphenated MS is increasingly being utilized for drug testing in place of IAs (Pasin et al., 2017). While there are just a few known chemicals covered by focused MS drug screening, many hundred analytes can be detected with the use of contemporary multi-analyte LC-MS/MS techniques with selective reaction monitoring (Mbughuni et al., 2016). Naturally, the number of monitored transitions and selectivity both affect identification power (Peters, 2011; Panderi et al., 2017). The powerful search algorithms and well-developed reference libraries including over 10,000 selective electron impact spectra, GC-MS remains a viable option for comprehensive drug screening, which is primarily conducted on urine (Meyer et al., 2010; Maurer et al., 2007; Grapp et al., 2016). Nevertheless, it was determined that LC-MSn screening using a matching reference library was an appropriate addition to GC-MS (Wissenbach et al.,

2011-193]. While this was going on, targeted and thorough screening was also successfully carried out using LC-HRMS/MS with TOF or OT analyzers. This method offered several benefits, including exceptionally strong identifying capability with relatively simple method development, great selectivity, sensitivity, robustness, and flexibility (Maurer and Meyer 2016; Beck and Ericsson, 2014; Concheiro et al., 2015). The advantages and disadvantages of low-resolution mass spectrometry (LRMS) and high-resolution mass spectrometry (HRMS) for NPS screening in hair, blood, urine, and soil that has been soaked with urine gathered throughout rave events have already been covered by Meyer and Maurer (Meyer and Maurer, 2016) elsewhere. Although most writers still create targeted screening processes, they concluded that HRMS could handle non-targeted screening operations (generic unknown screening) more easily. Additionally, they talked about the benefits of using extremely rapid scanning (QTOF) devices for sequential window acquisition of all theoretical fragment ion spectra (SWATH), which is a promising method for non-targeted HRMS screening in clinical and forensic toxicology (Roemmelt et al., 2015; Karastogianni et al., 2017). The first HRMS reference library was created and effectively used for urine screening, concentrating on metabolites (Maurer, 2017). Metabolite detection improves selectivity, permits confirmation of body passage, and, ultimately, reduces the possibility of false-negative LC-MS results that can be brought on by ion suppression of the target analyte (George et al., 2018; Xu, 2016). Examining metabolic trends can even lower the chance of false positive outcomes. Understanding these patterns is necessary to create screening methods based on metabolites. If the metabolites pharmacologic and/or toxic effects, these studies are also crucial for toxicological risk assessment in development. drug discovery and controlled human studies are prohibited, the flood of non-preclinical substances (NPS) has encouraged toxicologists to conduct metabolism studies in animals or human liver preparations, such as primary hepatocytes, cell cultures, S9 fractions, microsomes, or cytosol. NPS are sold and consumed without any preclinical testing (Maurer and Meyer, 2016, Beck and Ericsson, 2014) . Richter et al., (2017) examined NPS metabolism data from research involving human urine, primary human hepatocytes, and liver preparations. They came to the conclusion that human liver preparations, especially the pooled S9 fraction, were a sufficient and more affordable alternative in the context of metabolic investigations and for creating toxicological urine tests, even though hepatocytes offered the widest diversity of metabolites. Because

of its exceptional sensitivity and identifying power, HRMS is the industry leader in this area [Maurer and Meyer, 2016, Maurer and Meyer, 2016, Maurer and Meyer, 2012). It also plays a significant role in research on drug transporters or drug binding to proteins (Meyer, 2016 Meyer, 2015 Mardal et al., 2016). Lastly, the high sensitivity of contemporary equipment opens up new research and application domains, such as the miniaturization of drug (of abuse) quantification using dried blood spots (Stove et al., 2012; Verplaetse and Henion, Patteet et al., 2015; Berm et al., 2015; Ambach et al., 2014) or the detection of chemical warfare agent protein adducts

as a biomarker of poisoning in innovative micro sampling devices (John et al., 2016). Successfully applied to a dried urine spot LC-MSⁿ screening are well-established comprehensive urine screening methodologies (Michely, et al., 2017). Of course, some of its drawbacks had to be mentioned, such the fact that some very low-dose medications lacked adequate detection limits. Additionally, this dried urine spot screening was moved to an HRMS without chromatography or extraction. Urine could be directly screened for drugs (of abuse) and their metabolites using paper-spray ionization (Michely et al., 2017).

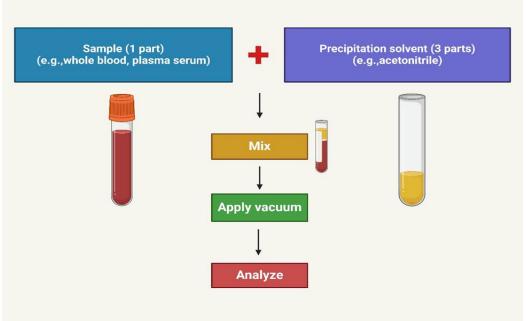


Fig. 2. An investigation of the process of preparing samples in the field of forensic toxicology

6. Clinical toxicology

clinical emergency toxicology, sample transportation, laboratory analysis, and findings reporting are allotted no more than two hours. Analytical techniques that can be applied in this situation are strictly required by the time constraint. The primary use of urine is in drug testing. Immunoassay methods are widely used in hospital laboratories due to their ease of use and speed. However, it is generally recognized that the present generation of immunoassay techniques can only cover a small portion of compounds that are significant to toxicology. Gamma-hydroxybutyrate, the majority of designer pharmaceuticals, oral antidiabetics, calcium channel blockers, β-blockers, and pregabalin are only a few of the numerous significant ones that are overlooked. A comparative analysis between immunoassay and full GC-MS screening revealed that nearly every other patient had consumed chemicals that the immunoassay had not been able to identify. Furthermore, the results of the GC-MS did not agree with every fifth result

(von Mach, et al., 2007). According to certain reports, the majority of acute poisoning patients would not respond well to treatment if substances taken in overdose were quickly identified through thorough drug screening (Pohjola-Sintonen et al., 2000). The same authors did note, nevertheless, that thorough screening would allow for the best possible care for many acute poisoning cases by eliminating the need for expensive medications and supervision and making it easier to identify instances that would need immediate drug-specific treatment (Pohjola-Sintonen et al., 2000). The program has generally been favorably regarded in the centers where comprehensive broad-spectrum drug screening has been organized appropriately. Another problem is that without the toxicologist's assistance and direction, doctors may not always be able to understand the analysis's findings (Hammett -Stabler et al., 2002; Tenore 2010; Flanagan, 2004).

7. Forensic toxicology

There are many different areas in forensic toxicology (Table 1), and each has different requirements for the breadth and depth of screening. Comprehensive screening is ideal in the following areas: drug-facilitated crime; child welfare; driving while intoxicated; post-mortem toxicology; and assault victim and offender investigations. Testing of drug in the penal system, the workplace, and the military forces are among the settings focused on drugs of abuse (Vuori and Ojanperä, 2009). The same kinds of specimens used in clinical toxicology are also used in clinical forensic toxicology. But unlike clinical toxicology, the forensic environment places more value on analytical findings that can be supported in court. As such, a confirmation step ought to be conducted in conjunction with screening in forensic toxicology, either concurrently with the same analytical run or independently. Considering that

someone who tests positive for an unlawful drug may face consequences, the treatment of drugdependent individuals falls between the purview of clinical and forensic toxicology. It is standard practice in post-mortem toxicology to use two distinct specimens and two distinct procedures. Because of the varied and frequently deteriorated character of the specimens as well as the wide variety of specimens accessible for investigation, applying analytical techniques in postmortem toxicology is frequently more challenging than in other forms of forensic toxicology (Drummer et al., 2007). It is important to thoroughly validate methods for the specific postmortem specimen being used. In addition to blood, urine and vitreous humour can be useful specimens. In certain situations, solid tissues like the liver and the contents of the stomach can also be used (Drummer et al., 2007).

Table 1. An overview of the features of a few time-of-flight and Orbitrap Fourier-transform mass spectrometry-based drug-screening techniques for doping control and clinical and forensic

toxicology, arranged chronologically by year of publication

toxicology, arranged chronologically by year of publication.						
Analytes	Work-up	Matrix	HRMS technique	ID procedure	Qual/quant	Ref.
637 drugs and metabolites	Mixed mode SPE	Urine	LC-TOFMS	Am, RT, minimum abundance criteria, and precursor exact mass reverse database search and EIC generation	Qual	(Pelander et al., 2003)
97 doping agents	Mixed mode SPE	Urine	LC-TOFMS	Minimum abundance criteria, Δm, RT, isotope pattern matching, and precursor exact mass reverse database search	Qual	(Kolmonen et al., 2007)
29 doping agents	LLE	Urine	LC– OrbitrapMS	Find the precise masses of the precursor and diagnostic fragments; indicate any unreported criteria.	Qual	(Virus et al., 2008)
10 drugs of abuse and metabolites	LLE	Hair	CE_TOFMS	Generate elemental composition for precursors within specified RT windows, match isotopic patterns, and meet Δm requirements	Qual and semiquant	(Gottardo et al., 2007)
175 drugs	Mixed mode SPE	Whole blood	UHPLC– TOFMS	Primary: Δm , RT, and minimum abundance parameters for searching the target database; additional: matching isotope patterns	Qual	(Dalsgaard et al., 2012)
815 drugs and metabolites	Mixed mode SPE	Hair	LC-TOFMS	Minimum abundance criteria, Δm, RT, isotope pattern matching, and precursor exact mass reverse database search	Qual	(Pelander et al., 2008)
815 drugs and metabolites	Mixed mode SPE	Vitreous humour	LC-TOFMS	Isotopic pattern, precursor precise mass reverse database search, Δm , and RT	Qual	(Pelander et al., 2010)
13 steroids	Dilution	Hormone preparations	LC-TOFMS and LC- QTOFM	Manual: precise mass-based elemental composition of predecessors TOFMS and QTOFMS selected from TIC, UV spectrum DBE criteria, fragment analysis by DDA or ISCID	Qual	(Nielen et al., 2001)

8. The present challenges in forensic and analytical toxicology

The main concerns concern behavioral or human performance toxicology, including drug testing in workplace, abstinence control, postmortem toxicology, impaired driving evaluation, and drugfacilitated crimes (Wyman, 2012). Within living organisms, mostly blood or urine, forensic toxicology primarily involves the quantitative and qualitative examination of poisons, prescription medicines, drugs of abuse (DOA), or ethanol, as

well as the interpretation of the corresponding data. Prescreen immunoassays (IA) are frequently used in routine laboratory procedures to screen for the most pertinent DOAs. These IA tests are frequently followed by confirmatory analyses hyphenated chromatographic methods, like liquid chromatography (LC) -MS (mass spectrometry) or gas chromatography (GC) —MS (Maurer, 2007; Drummer, 2007; Maurer, 2010). As of December 2017, over 800 NPS had been informed to the UNODC Early Warning Advisory, making the use

and abuse of these substances an international issue (United Nations Office on Drugs and Crime, 2018). Of these, the majority (68%) were stimulants and synthetic cannabinoids, which accounted for the majority of newly reported NPS in 2017. In general, there is a dearth of knowledge regarding the harmful impacts and toxicity of NPS, which is becoming a global issue. Furthermore, because of their transience on the drug scene, direct detection and identification continue to be difficult for analysis. Since most of the time common IAs cannot accurately identify entire classes of NPS, thorough screening procedures must be developed to detect them. Although highly sensitive, focused techniques such as multiple reaction monitoring (MRM) require frequent updates and reference standards that are either hard to come by or very expensive. Due to its ability to evaluate data retrospectively and eliminate the requirement for technique adjustment, HRMS has demonstrated great promise (Grabenauer et al., 2012; Shanks et al., 2012). Developing innovative screening techniques that do not specifically target the chemical structures of the analyte or its metabolites is an alternate strategy. This could be quite helpful in providing prompt action in the event of suspected NPS intake and in helping to resolve this intricate analytical problem. The first method demonstrates that an assay that can identify synthetic cannabinoids and their metabolites according to their interaction and activity with cannabinoid receptors may be developed (Cannaert et al., 2017). As a primary urine screening tool, such an activity-based screening test may supplement traditional analytical techniques (both targeted and untargeted). In contrast to targeted techniques, which miss compounds if they are not on the candidate list, this method may result in fewer false negative results even though it is hard to positively identify individual synthetic cannabinoids (Bijlsma et al., 2018). Drug users may turn to NPS not only to obtain legal highs but also, possibly, to avoid testing positive for drugs on drug screening tests. In situations when drug abstinence must be demonstrated, such as driving liability tests, certain psychiatric or jail settings, or workplace pharmacological screening processes, the latter is especially pertinent (Bijlsma et al., 2018). Urine continues to be the preferred matrix in settings for abstinence control (Verstraete, 2004; Phan et al., 2012; Fu et al., 2014). Therefore, labs must identify attempts at urine adulteration that may be made in an attempt to evade positive drug test findings (Wu et al., 1999). Diluting real pee, replacing it with synthetic urine, or chemical adulteration are common manipulation techniques. Because several compounds are known to hide drug detection effects, their use has been documented. Everyday domestic substances like pyridinum chlorochromate

(PCC), hypochlorite-based bleach (NaOCl), peroxide (H₂O₂), and peroxidase are often appropriate for chemical urine adulteration (Fu et al., 2014; Uebel and Wium, 2002; Jaffee et al., 2007). Products like Stealth® (consisting of H2O2 and peroxidase) (Valtier and Cody, 2002) Klear® (consisting of KNO2) (Peace and Tarnai, 2002), or Whizzies® (consisting of sodium nitrite) (Dasgupta et al., 2004) and "Urine Luck" (consisting of PCC) (Wu et al., 1999; Paul et al., 2000) are easily accessible online (Fu et al., 2014; Jaffee et al., Dasgupta, 2007). Products designed to adulterate urine for commercial use are in particular common in the United States. This also holds for artificial urine products that are sold commercially (Goggin et al., 2017; Kluge et al., 2018). Comprehensive testing is sometimes impeded by time, costs, and resources, even though screening for a wide range of chemical adulterants and artificial urine products be a requirement for toxicological laboratories. Integrity testing, integrated sample checks, and spot and dipstick tests are quicker and less expensive alternatives to IA systems that are sold commercially. However, these are frequently linked to high percentages of false negative or inaccurate positive outcomes (Fu et al., 2014, Edwards et al., 1993, Matriciani et al., 2018).

9. Conclusions

TDM is crucial in clinical and forensic toxicology as it aids in optimizing drug therapy and identifying drug-related incidents. TDM is a crucial tool in clinical and forensic toxicology, providing valuable insights into drug therapy management and incident investigation. Despite challenges like metabolism variability, TDM optimizes treatment, minimizes adverse effects, and enhances forensic analyses. It reveals that despite providing valuable insights, faces significant challenges TDM also limitations. The widespread adoption effectiveness of drug metabolism tests in clinical practice are delayed by variability in metabolism, assay inaccuracies, and interpretational complexities. investigations Forensic face challenges like postmortem redistribution and analytical complicating sensitivities, the interpretation of TDM results and requiring careful integration with comprehensive forensic analyses. Future efforts should focus on addressing challenges and enhancing the utility and reliability of TDM in clinical and forensic conditions. Analytical techniques advancements, including the creation of more sensitive and specific assays, are crucial for enhancing the accuracy and precision of drug measurements. Interdisciplinary collaboration between pharmacologists, toxicologists, forensic scientists, and clinicians is crucial for developing standardized protocols and guidelines for TDM

interpretation and integration into clinical and practice. Advancements pharmacogenomics and personalized medicine provide opportunities to customize drug therapy on individual patient characteristics, enhancing therapeutic outcomes and minimizing adverse effects. Future research should address TDM challenges by improving assay sensitivity, standardizing interpretation guidelines, adopting LC-MS for accuracy, and integrating pharmacogenomic information for personalized medicine approaches. Emerging technologies like LC-MS and point-of-care testing can enable rapid and reliable drug monitoring in various clinical and conditions. TDM, despite challenges, holds great promise for improving patient care, enhancing drug safety, and facilitating toxicology forensic investigations. Point-of-care testing and mobile technologies can improve clinical drug monitoring, while collaboration between forensic scientists and clinicians is crucial for standardized protocols and comprehensive forensic analyses. TDM's future lies in embracing innovations, technological interdisciplinary collaboration, and refining methodologies to enhance patient care and facilitate toxicology forensic investigations.

Reference

- Abernethy, D. R., Greenblatt, D. J., & Shader, R. I. (1985). Imipramine and desipramine disposition in the elderly. Journal of Pharmacology and Experimental Therapeutics, 232(1), 183-188.
- 2. Adityanjee, A. J., & Srivastava, A. S. (2006). Clinical pharmacokinetics of antipsychotics in children and adolescents: a critical review of the literature. Clinical Pharmacokinetics, 45(11), 1059-1084.
- 3. Aichhorn, W., Marksteiner, J., Walch, T., Zernig, G., & Saria, A. (2006). Pramipexole in treatment-resistant depression: a 16-week naturalistic study. European Neuropsychopharmacology, 16(6), 464-471.
- Hadithy, A. F., Ivanova, S. A., Pechlivanoglou, P., Semke, A., Fedorenko, O., Kornetova, E., Ryadovaya, L., Brouwers, J. R., Wilffert, B., Bruggeman, R., & Loonen, A. J. (2009). Tardive dyskinesia and DRD3, HTR2A and HTR2C gene polymorphisms in Russian psychiatric inpatients from Siberia. Progress in neuropsychopharmacology biological & psychiatry, 33(3), 475-481.
- Al-Janabi, I., Arranz, M. J., Blakemore, A. I., Saiz, P. A., Susce, M. T., Glaser, P. E., Clark, D., & de Leon, J. (2009). Association study of serotonergic gene variants with antipsychotic-

- induced adverse reactions. Psychiatric genetics, 19(6), 305–311. https://doi.org/10.1097/YPG.0b013e328332
- Ambach, L., Hernández Redondo, A., König, S., & Weinmann, W. (2014). Rapid and simple LC-MS/MS screening of 64 novel psychoactive substances using dried blood spots. Drug testing and analysis, 6(4), 367-375.
- 7. Armijo, J. A., Cuadrado, A., Bravo, J., & Arteaga, R. (1997). Vigabatrin serum concentration to dosage ratio: influence of age and associated antiepileptic drugs. Therapeutic drug monitoring, 19(5), 491–498. https://doi.org/10.1097/00007691-1997100 00-00001.
- 8. Aronson, J. K., & Ferner, R. E. (2016). The law of mass action and the pharmacological concentration—effect curve: resolving the paradox of apparently non-dose-related adverse drug reactions. British Journal of Clinical Pharmacology, 81(1), 56-61.
- 9. Åsberg, M., Crönholm, B., Sjöqvist, F., & Tuck, D. (1971). Relationship between plasma level and therapeutic effect of nortriptyline. British Medical Journal, 3(5770), 331-334.
- Azad, A. K., Praveen, M., & Sulaiman, W. M. A. B. W. (2024). Assessment of Anticancer Properties of Plumbago zeylanica. Harnessing Medicinal Plants in Cancer Prevention and Treatment, 91–121.
- 11. https://doi.org/10.4018/979-8-3693-1646-7.ch004
- 12. Backman, J. T., Filppula, A. M., Niemi, M., & Neuvonen, P. J. (2016). Role of cytochrome P450 2C8 in drug metabolism and interactions. Pharmacological Reviews, 68(1), 168-241.
- 13. Balant, L., Balant-Gorgia, A., Eisele, R., Gex-Fabry, M., & Garrone, G. (1989). Clinical and pharmacokinetic evaluation of zuclopenthixol acetate in Viscoleo®. Pharmacopsychiatry, 22(06), 250-254.
- Baranczewski, P., Stanczak, A., Sundberg, K., Svensson, R., Wallin, Å., Jansson, J., & Edlund, P. O. (2006). Introduction to in vitro estimation of metabolic drug–drug interactions: basics and principles. Chemico-Biological Interactions, 164(1-2), 187-206.
- 15. Barcelo, B., Noce, V., & Gomila, I. (2018). Building bridges between clinical and forensic toxicology laboratories. Current Pharmaceutical Biotechnology, 19(2), 99-112.
- Barr, J. T., Rodriguez-Cruz, V., Smith, C., & Muszynski, M. (2002). Pharmacokinetic considerations in psychiatric research. In H. M. Kranzler & D. W. Alderson (Eds.), Research methods in psychiatry (pp. 195-211). American Psychiatric Association Publishing.

17. Barski, O. A., Tipparaju, S. M., & Bhatnagar, A. (2008). The aldo-keto reductase superfamily and its role in drug metabolism and detoxification. Drug Metabolism Reviews, 40(4), 553-624.

- 18. Bauer, L. A. (2008). Applied clinical pharmacokinetics. McGraw-Hill Medical New York.
- 19. Baumann, P., Hiemke, C., Ulrich, S., Eckermann, G., Gaertner, I., Gerlach, M., Kuss, H. J., Laux, G., Müller-Oerlinghausen, B., Rao, M. L., Riederer, P., Zernig, G., & Arbeitsge-meinschaft fur neuropsychopharmakologie und pharmakopsychiatrie (2004). The AGNP-TDM expert group consensus guidelines: therapeutic drug monitoring in psychiatry. Pharmacopsychiatry, 37(6), 243–265. https://doi.org/10.1055/s-2004-832687
- Baumann, P., Kirchherr, H., Berney, P., & Hiemke, C. (2012). Flupentixol: relevance of stereoselective therapeutic drug monitoring. Psychopharmacology, 221(4), 719-720.
- 21. Baumann, P., Zullino, D. F., & Eap, C. B. (2002). Enantiomers' potential in psychopharmacology—a critical analysis with special emphasis on the antidepressant escitalopram. European Neuropsychopharmacology, 12(5), 433-444.
- 22. Beedham, C., Miceli, J. J., & Obach, R. S. (2003). Ziprasidone metabolism, aldehyde oxidase, and clinical implications. Journal of Clinical Psychopharmacology, 23(3), 229-232.
- 23. Benedetti, M. S., Whomsley, R., Baltes, E., Tonner, F., Muller, L., & Kleinermans, D. (2009). Drug metabolism and pharmacokinetics. Drug Metabolism Reviews, 41(3), 344-390.
- 24. Berm, E. J., Paardekooper, J., Brummel-Mulder, E., Hak, E., Wilffert, B., & Maring, J. G. (2015). A simple dried blood spot method for therapeutic drug monitoring of the tricyclic antidepressants amitriptyline, nortriptyline, imipramine, clomipramine, and their active metabolites using LC-MS/MS. Talanta, 134, 165-172.
- Bertelsen, K. M., Venkatakrishnan, K., Von Moltke, L. L., Obach, R. S., & Greenblatt, D. J. (2003). Apparent mechanism-based inhibition of human CYP2D6 in vitro by paroxetine: comparison with fluoxetine and quinidine. Drug Metabolism and Disposition, 31(3), 289-293.
- 26. Biernacka, J., et al., (2015). The International SSRI Pharmacogenomics Consortium (ISPC): a genome-wide association study of antidepressant treatment response. Translational Psychiatry, 5(4), e553.

27. Bijlsma, L., Gil-Solsona, R., Hernández, F., & Sancho, J. V. (2018). What about the herb? A new metabolomics approach for synthetic cannabinoid drug testing. Analytical and bioanalytical chemistry, 410, 5107-5112.

- Birkenhäger, T. K., Moleman, P., Boonstra, H., & Schene, A. H. (2006). Lack of evidence for the efficacy of antipsychotics in treatmentresistant depression: a meta-analysis. Journal of Clinical Psychopharmacology, 26(6), 650-654.
- 29. Blake, C. M., & Ridout, G. (2007). Antidepressants and cytochrome P450 2D6 (CYP2D6) inhibition: are there clinically relevant drug interactions? Canadian Journal of Psychiatry, 52(11), 780-788.
- 30. Breyer-Pfaff, U., & Nill, K. (2004). Carbonyl reduction of naltrexone and dolasetron by oxidoreductases isolated from human liver cytosol. Journal of Pharmacy and Pharmacology, 56(12), 1601-1606.
- 31. Bruijn, J. A., Moleman, P., Mulder, P. G., van den Broek, W. W., van Hulst, A. M., van der Mast, R. C., & van de Wetering, B. J. (1996). A double-blind, fixed blood-level study comparing mirtazapine with imipramine in depressed inpatients. Psychopharmacology, 127(3), 231–237.
- 33. Buko, A. (2017). Capillary electrophoresis mass spectrometry based metabolomics. Journal of Applied Bioanalysis, 3(1), 5–20.
- 34. Bymaster, F. P., Calligaro, D. O., Falcone, J. F., Marsh, R. D., Moore, N. A., Tye, N. C., ... & Wong, D. T. (1996). Radioreceptor binding profile of the atypical antipsychotic olanzapine. Neuropsychopharmacology, 14(2), 87-96.
- Callaghan, J. T., Bergstrom, R. F., Ptak, L. R.,
 Beasley, C. M. (1999). Olanzapine: pharmacokinetic and pharmacodynamic profile. Clinical Pharmacokinetics, 37(3), 177-193.
- 36. Cannaert, A., Franz, F., Auwärter, V., & Stove, C. P. (2017). Activity-based detection of consumption of synthetic cannabinoids in authentic urine samples using a stable cannabinoid reporter system. Analytical chemistry, 89(17), 9527-9536.

37. Castberg, I., Skogvoll, E., & Spigset, O. (2007). Quetiapine and drug interactions: evidence from a routine therapeutic drug monitoring service. Journal of Clinical Psychiatry, 68(10), 1540-1545.

- 38. Chenu, F., Batten, L. A., Zernig, G., Ladstaetter, E., Hébert, C., & Blier, P. (2009). Comparison of pharmacokinetic profiles of brand-name and generic formulations of citalopram and venlafaxine: a crossover study. The Journal of clinical psychiatry, 70(7), 958–966.
 - https://doi.org/10.4088/jcp.09m05315.
- 39. Chung, H., & Choe, S. (2017). Overview of forensic toxicology, yesterday, today and in the future. Current Pharmaceutical Design, 23(36), 5429-5436.
- 40. Clark, R. (2011). Therapeutic drug monitoring in psychiatric practice: clinical implications. Psychiatric Times, 28(3), 35-37.
- 41. Concheiro, M., Castaneto, M., Kronstrand, R., & Huestis, M. A. (2015). Simultaneous determination of 40 novel psychoactive stimulants in urine by liquid chromatographyhigh resolution mass spectrometry and library matching. Journal of chromatography. A, 1397, 32–42.
- 42. Court, M. H. (2010). Interindividual variability in hepatic drug glucuronidation: studies into the role of age, sex, enzyme inducers, and genetic polymorphism using the human liver bank as a model system. Drug Metabolism Reviews, 42(1), 209-224.
- 43. Dalsgaard, P. W., Rasmussen, B. S., Müller, I. B., & Linnet, K. (2012). Toxicological screening of basic drugs in whole blood using UPLC-TOF-MS. Drug Testing and Analysis, 4(5), 313-319.
- 44. Dasgupta, A. (2007). The effects of adulterants and selected ingested compounds on drugs-of-abuse testing in urine. American Journal of Clinical Pathology, 128(3), 491-503.
- 45. Dasgupta, A., Chughtai, O., Hannah, C., Davis, B., & Wells, A. (2004). Comparison of spot tests with AdultaCheck 6 and Intect 7 urine test strips for detecting the presence of adulterants in urine specimens. Clinica Chimica Acta, 348(1-2), 19-25.
- 46. Dawling, S. (1982). Monitoring of tricyclic antidepressant therapy. Clinical Biochemistry, 15(1), 56-61.
- 47. de Leon, J. (2003). The effect of atypical versus typical antipsychotics on tardive dyskinesia: a naturalistic study. European Archives of Psychiatry and Clinical Neuroscience, 253(2), 103-107.
- 48. de Leon, J. (2014). Evidence-based medicine versus personalized medicine: are they

enemies? Journal of Clinical Psychopharmacology, 34(2), 153-160.

- 49. de Leon, J., Armstrong, S. C., Cozza, K. L., & Clinical Practice, L. O. S. A. (2006). The dosing of atypical antipsychotics. Psychosomatics, 47(3), 277-283.
- 50. de Leon, J., Spina, E., & Diaz, F. J. (2013). Clobazam therapeutic drug monitoring: a comprehensive review of the literature with proposals to improve future studies. Therapeutic drug monitoring, 35(1), 30-47.
- 51. de Leon, J., Susce, M. T., & Murray-Carmichael, E. (2006). The AmpliChip CYP450 genotyping test: Integrating a new clinical tool into clinical practice. Molecular Diagnosis & Therapy, 10(3), 135-151.
- 52. Diaz, F. J., Santoro, V., Spina, E., Cogollo, M., Rivera, T. E., Botts, S., & de Leon, J. (2008). Estimating the size of the effects of comedications on plasma clozapine concentrations using a model that controls for clozapine doses and confounding variables. Pharmacopsychiatry, 41(3), 81–91. https://doi.org/10.1055/s-2007-1004591.
- 53. Dinis-Oliveira, R. J. (2016). Metabolomics of methadone: clinical and forensic toxicological implications and variability of dose response. Drug Metabolism Reviews, 48(4), 568-576.
- 54. Domschke, K., Tidow, N., Schwarte, K., Deckert, J., Lesch, K. P., Arolt, V., Zwanzger, P., & Baune, B. T. (2014). Serotonin transporter gene hypomethylation predicts impaired antidepressant treatment response. The international journal of neuropsychopharmacology, 17(8), 1167–1176. https://doi.org/10.1017/S146114571400039X
- 55. Dost, F. (1953). Kinetik de Konzentrationsabläufe in der Kreislaufflüssigkeit. Leipzig: Thieme.
- 56. Dresser, G. K., Spence, J. D., & Bailey, D. G. (2000). Pharmacokinetic–pharmacodynamic consequences and clinical relevance of cytochrome P450 3A4 inhibition. Clinical Pharmacokinetics, 38(1), 41-57.
- 57. Drummer, O. H. (2007). Requirements for bioanalytical procedures in postmortem toxicology. Analytical and bioanalytical chemistry, 388, 1495-1503.
- 58. Eap, C. B., Bender, S., & Baumann, P. (2000). Newer antidepressants: pharmacokinetics, therapeutic drug monitoring and pharmacogenetics. Therapeutic Drug Monitoring, 22(1), 137-143.
- 59. Eap, C. B., Buclin, T., & Baumann, P. (2002). Interindividual variability of the clinical pharmacokinetics of methadone: implications for the treatment of opioid dependence. Clinical Pharmacokinetics, 41, 1153-1193.

60. Edwards, C., Fyfe, M. J., Liu, R. H., & Walia, A. (1993). Evaluation of common urine specimen adulteration indicators. Journal of analytical toxicology, 17(4), 251-252.

- 61. Egberts, K., Mehler-Wex, C., & Gerlach, M. (2011). Therapeutic drug monitoring in child and adolescent psychiatry. Pharmacopsychiatry, 21(06), 249-253.
- 62. Ellingrod, V. L., Perry, P. J., & Lauriello, J. (2003). Genetic polymorphisms and antipsychotic metabolism. American Journal of Medical Genetics Part B: Neuropsychiatric Genetics, 123B(1), 50-58.
- 63. Ereshefsky, L. (2009). Pharmacokinetics and drug interactions: update for new antipsychotics. Journal of Clinical Psychiatry, 70(4), 27-33.
- 64. European Monitoring Centre for Drugs and Drug Addiction. (2009). Understanding the "Spice" phenomenon.
- 65. Evans, W. E., & Relling, M. V. (1999). Pharmacogenomics: translating functional genomics into rational therapeutics. Science, 286(5439), 487-491.
- 66. Farde, L., Nordström, A. L., & Wiesel, F. A. (1992). Positron emission tomographic analysis of central D1 and D2 dopamine receptor occupancy in patients treated with classical neuroleptics and clozapine: relation to extrapyramidal side effects. Archives of General Psychiatry, 49(7), 538-544.
- 67. Ferrell, P. B., & McLeod, H. L. (2008). Carbamazepine, HLA-B 1502 and risk of Stevens–Johnson syndrome and toxic epidermal necrolysis: US FDA recommendations.
- 68. Flanagan, R. J. (2004). Developing an analytical toxicology service: principles and guidance. Toxicological reviews, 23, 251-263.
- 69. Fleischhacker, W. W., Meise, U., Gunther, V., Kurz, M., & Deix, K. (1994). Serum concentrations of clozapine and norclozapine in patients with schizophrenia on concomitant treatment. Journal of Clinical Psychopharmacology, 14(2), 128-132.
- 70. Flockhart, D. A., Oesterheld, J. R., & Gandolfi, A. J. (2000). Pharmacogenetic principles and clinical applications. Mayo Clinic Proceedings, 75(7), 747-754.
- 71. Fu, S., Luong, S., Pham, A., Charlton, N., & Kuzhiumparambil, U. (2014). Bioanalysis of urine samples after manipulation by oxidizing chemicals: technical considerations. Bioanalysis, 6(11), 1543-1561.
- 72. Garg, U., & Dasouki, M. (2006). Expanded newborn screening of inherited metabolic disorders by tandem mass spectrometry:

- clinical and laboratory aspects. Clinical biochemistry, 39(4), 315-332.
- 73. Garg, U., & Zhang, Y. V. (2016). Mass spectrometry in clinical laboratory: applications in therapeutic drug monitoring and toxicology. In Clinical applications of mass spectrometry in drug analysis: methods and protocols (pp. 1-10).
- 75. Gerlach, M., Egberts, K., Dang, S. Y., Plener, P., Taurines, R., Mehler-Wex, C., & Romanos, M. (2016). Therapeutic drug monitoring as a measure of proactive pharmacovigilance in child and adolescent psychiatry. Expert opinion on drug safety, 15(11), 1477–1482. https://doi.org/10.1080/14740338.2016.122 5721
- 76. Gervasini, G., Carrillo, J. A., & Benitez, J. (2004). Potential role of cerebral cytochrome P450 in clinical pharmacokinetics: modulation by endogenous compounds. Clinical Pharmacokinetics, 43, 693-706.
- 77. Gex-Fabry, M., Balant-Gorgia, A. E., & Balant, L. P. (2003). Therapeutic drug monitoring of olanzapine: the combined effect of age, gender, smoking, and comedication. Therapeutic drug monitoring, 25(1), 46–53. https://doi.org/10.1097/00007691-200302000-00007
- 78. Ghosh, C., Marchi, N., Desai, N., Puvenna, V., Hossain, M., Gonzalez-Martinez, J., & Janigro, D. (2011). Cellular localization and functional significance of CYP3A4 in the human epileptic brain. Epilepsia, 52(3), 562-571.
- 79. Goggin, M. M., Tann, C. M., Miller, A., Nguyen, A., & Janis, G. C. (2017). Catching fakes: new markers of urine sample validity and invalidity. Journal of Analytical Toxicology, 41(2), 121-126.
- 80. González, M. M., Akamine, Y., & Shimosegawa, E. (2015). Considerations on the pharmacogenetics of psychiatric drugs. Drug Metabolism and Pharmacokinetics, 30(1), 1-9.
- 81. Gottardo, R., Fanigliulo, A., Bortolotti, F., De Paoli, G., Pascali, J. P., & Tagliaro, F. (2007). Broad-spectrum toxicological analysis of hair based on capillary zone electrophoresis—time-

of-flight mass spectrometry. Journal of Chromatography A, 1159(1-2), 190-197.

- 82. Grabenauer, M., Krol, W. L., Wiley, J. L., & Thomas, B. F. (2012). Analysis of synthetic cannabinoids using high-resolution mass spectrometry and mass defect filtering: implications for nontargeted screening of designer drugs. Analytical chemistry, 84(13), 5574-5581.
- 83. Grapp, M., Maurer, H. H., & Desel, H. (2016). Systematic forensic toxicological analysis by GC-MS in serum using automated mass spectral deconvolution and identification system. Drug testing and analysis, 8(8), 816-825.
- 84. Greenbaum, L., Smith, R. C., Rigbi, A., Strous, R., Teltsh, O., Kanyas, K., Korner, M., Lancet, D., Ben-Asher, E., & Lerer, B. (2009). Further evidence for association of the RGS2 gene with antipsychotic-induced parkinsonism: protective role of a functional polymorphism in the 3'-untranslated region. The pharmacogenomics journal, 9(2), 103–110. https://doi.org/10.1038/tpj.2008.6
- 85. Greenbaum, L., Strous, R. D., Kanyas, K., Merbl, Y., Horowitz, A., Karni, O., Katz, E., Kotler, M., Olender, T., Deshpande, S. N., Lancet, D., Ben-Asher, E., & Lerer, B. (2007). Association of the RGS2 gene extrapyramidal symptoms induced by treatment with antipsychotic medication. Pharmacogenetics and genomics, 17(7), 519-528. https://doi.org/10.1097/FPC.0b013e32800ff
- 86. Gressier, F., Porcelli, S., Calati, R., & Serretti, A. (2016). Pharmacogenetics of clozapine response and induced weight gain: A comprehensive review and meta-analysis. European Neuropsychopharmacology, 26(2), 163-185.
- 87. Gupta, S. K., Shah, J. C., & Hwang, S. S. (1999). Pharmacokinetic and pharmacodynamic characterization of OROS® and immediate-release amitriptyline. British Journal of Clinical Pharmacology, 48(1), 71-78.
- 88. Haddad, P. M., & Sharma, S. G. (2007). Adverse effects of atypical antipsychotics: differential risk and clinical implications. CNS Drugs, 21(11), 911-936.
- 89. Haen, E. (2011). Therapeutic drug monitoring in pharmacovigilance and pharmacotherapy safety. Pharmacopsychiatry, 21(06), 254-258.
- 90. Haen, E., Greiner, C., Bader, W., & Wittmann, M. (2008). Expanding therapeutic reference ranges using dose-related reference ranges. Der Nervenarzt, 79, 558-566.

91. Hammett-Stabler, C. A., Pesce, A. J., & Cannon, D. J. (2002). Urine drug screening in the medical setting. Clinica chimica acta, 315(1-2), 125-135.

- 92. Hanley, J. A., & McNeil, B. J. (1982). The meaning and use of the area under a receiver operating characteristic (ROC) curve. Radiology, 143(1), 29-36.
- 93. Hefner, G., Laib, A. K., Sigurdsson, H., Hohner, M., & Hiemke, C. (2013). The value of drug and metabolite concentration in blood as a biomarker of psychopharmacological therapy. International review of psychiatry (Abingdon, England), 25(5), 494–508. https://doi.org/10.3109/09540261.2013.836 475.
- 94. Hefner, G., Ludwig, R., Leweke, F. M., Mössner, R., & Rao, M. L. (2013). An update on therapeutic drug monitoring of antipsychotics in children and adolescents. European Neuropsychopharmacology, 23(10), 811-821.
- 95. Hefner, G., Mössner, R., Schimke, J., Schosser, A., Maier, W., Benninghoff, J., & Remschmidt, H. (2015). Melperone but not bisoprolol or metoprolol is a clinically relevant inhibitor of CYP2D6: evidence from a therapeutic drug monitoring survey. Journal of Neural Transmission, 122, 1609-1617.
- Hegerl, U., Bottlender, R., Gallinat, J., Kuss, H.-J., Ackenheil, M., & Möller, H.-J. (1998). The serotonin syndrome scale: first results on validity. European Archives of Psychiatry and Clinical Neuroscience, 248, 96-103.
- 97. Hendershot, C. S. (2014). Pharmacogenetic approaches in the treatment of alcohol use disorders: addressing clinical utility and implementation thresholds. Addiction Science & Clinical Practice, 9, 1-8.
- 98. Hiemke, C., Baumann, P., Bergemann, N., Conca, A., Dietmaier, O., Egberts, K., Fric, M., Gerlach, M., Greiner, C., Gründer, G., Haen, E., Havemann-Reinecke, U., Jaquenoud Sirot, E., Kirchherr, H., Laux, G., Lutz, U. C., Messer, T., Müller, M. J., Pfuhlmann, B., Rambeck, B., ... Zernig, G. (2011). AGNP Consensus Guidelines for Therapeutic Drug Monitoring in Psychiatry: Update 2011. Pharmacopsychiatry, 44(6), 195–235. https://doi.org/10.1055/s-0031-1286287
- 99. Hiemke, C., Bergemann, N., Clement, H. W., Conca, A., Deckert, J., Domschke, K., ... & Zernig, G. (2018). Consensus guidelines for therapeutic drug monitoring in neuropsychopharmacology: update 2017. Pharmacopsychiatry, 51(1-2), 9-62.
- 100. Ho, Y. P., & Reddy, P. M. (2011). Advances in mass spectrometry for the identification of

pathogens. Mass spectrometry reviews, 30(6), 1203-1224.

- 101. Ingelman-Sundberg, M. (2004). Pharmacogenetics of cytochrome P450 and its applications in drug therapy: the past, present and future. Trends in Pharmacological Sciences, 25(4), 193-200.
- 102. Jaffee, W. B., Trucco, E., Levy, S., & Weiss, R. D. (2007). Is this urine really negative? A systematic review of tampering methods in urine drug screening and testing. Journal of Substance Abuse Treatment, 33(1), 33-42.
- 103. Jerling, M., Bertilsson, L., & Sjöqvist, F. (1994). The use of therapeutic drug monitoring data to document kinetic drug interactions: an example with amitriptyline and nortriptyline. Therapeutic drug monitoring, 16(1), 1-12.
- 104. Jimenez, C. R., & Verheul, H. M. (2014). Mass spectrometry-based proteomics: from cancer biology to protein biomarkers, drug targets, and clinical applications. American Society of Clinical Oncology Educational Book, 34(1), e504-e510.
- 105. John, H., Willoh, S., Hörmann, P., Siegert, M., Vondran, A., & Thiermann, H. (2016). Procedures for analysis of dried plasma using microsampling devices to detect sulfur mustard-albumin adducts for verification of poisoning. Analytical chemistry, 88(17), 8787-8794.
- 106. Jones, P. M., & Bennett, M. J. (2002). The changing face of newborn screening: diagnosis of inborn errors of metabolism by tandem mass spectrometry. Clinica Chimica Acta, 324(1-2), 121-128.
- 107. K. Hiemke, C. J. G., Baumann, P., Eckermann, G., Ulrich, S., & S. Zernig, G. (2010). Therapeutic drug monitoring in neuropsychopharmacology: an update on clinical practice guidelines. European Neuropsychopharmacology, 20(9), 671-684.
- 108. K. Meltzer, H. Y. (2012). Mechanisms of action of antipsychotic drugs. In A. Breier, M. Olfson, & S. Greenstein (Eds.), Schizophrenia: New pharmacological approaches (pp. 77-88). Springer.
- 109. Kaddurah-Daouk, R., Kristal, B. S., & Weinshilboum, R. M. (2008). Metabolomics: a global biochemical approach to drug response and disease. Annual Review of Pharmacology and Toxicology, 48, 653-683.
- 110. Kalow, W. (2001). Pharmacogenetics: a historical perspective. Current Drug Metabolism, 2(1), 1-11.
- 111. Kane, J. M., Kishimoto, T., & Correll, C. U. (2013). Assessing the impact of patient adherence on clinical outcomes in

- schizophrenia. Schizophrenia Research, 159(1), 247-253.
- 112. Karastogianni, S., Deliyanni, E. A., & Girousi, S. (2017). Application of promising carbonaceous materials in electrochemical DNA sensing. Journal of Applied Bioanalysis, 3(4), 110–119.
- 113. Keck Jr, P. E., McElroy, S. L., Strakowski, S. M., West, S. A., Sax, K. W., Hawkins, J. M., & Bourne, M. L. (1998). 12-month outcome of patients with bipolar disorder following hospitalization for a manic or mixed episode. American Journal of Psychiatry, 155(5), 646-652.
- 114. Kennedy, M. (2010). Post-mortem drug concentrations. Internal Medicine Journal, 40(3), 183-187.
- 115. Ketha, H., & Garg, U. (2020). An introduction to clinical and forensic toxicology. In Toxicology cases for the clinical and forensic laboratory (pp. 3-6). Elsevier.
- 116. Kirchheiner, J., & Seeringer, A. (2007). Clinical implications of pharmacogenetics of cytochrome P450 drug metabolizing enzymes. Biochimica et Biophysica Acta (BBA)-Proteins and Proteomics, 1770(3), 489-494.
- 117. Klengel, T., & Binder, E. B. (2013). Gene× environment interactions in the prediction of response to antidepressant treatment. International Journal of Neuropsychopharmacology, 16(3), 701-711.
- 118. Klotz, U. (2009). Pharmacokinetics and drug metabolism in the elderly. Drug Metabolism Reviews, 41(2), 67-76.
- 119. Kluge, J., Rentzsch, L., Remane, D., Peters, F. T., & Wissenbach, D. K. (2018). Systematic investigations of novel validity parameters in urine drug testing and prevalence of urine adulteration in a two-year cohort. Drug testing and analysis, 10(10), 1536-1542.
- 120. Koelch, M., Pfalzer, A. K., Kliegl, K., Rothenhöfer, S., Ludolph, A. G., Fegert, J. M., Burger, R., Mehler-Wex, C., Stingl, J., Taurines, R., Egberts, K., & Gerlach, M. (2012). Therapeutic drug monitoring of children and adolescents treated with fluoxetine. Pharmacopsychiatry, 45(2), 72–76. https://doi.org/10.1055/s-0031-1291294.
- 121. Kolmonen, M., Leinonen, A., Pelander, A., & Ojanperä, I. (2007). A general screening method for doping agents in human urine by solid phase extraction and liquid chromatography/time-of-flight mass spectrometry. Analytica Chimica Acta, 585(1), 94-102.
- 122. Kugelberg, F. C., Druid, H., Carlsson, B., Ahlner, J., & Bengtsson, F. (2004). Postmortem redistribution of the enantiomers

of citalopram and its metabolites: an experimental study in rats. Journal of analytical toxicology, 28(8), 631–637. https://doi.org/10.1093/jat/28.8.631

- 123. Lagacé-Wiens, P. (2015). Matrix-assisted laser desorption/ionization time of flight mass spectrometry (MALDI-TOF/MS)-based identification of pathogens from positive blood culture bottles. In Sepsis: Diagnostic Methods and Protocols (pp. 47-55). Springer.
- 124. Lam, Y. W. F., Banerjee, A., & McDonald, G. B. (2004). Pharmacokinetic and pharmacodynamic interactions of azole antifungal agents and cyclosporine. Clinical Pharmacokinetics, 43(10), 763-795.
- 126. Lerer, B., Segman, R. H., Tan, E. C., Basile, V. S., Cavallaro, R., Aschauer, H. N., Strous, R., Chong, S. A., Heresco-Levy, U., Verga, M., Scharfetter, J., Meltzer, H. Y., Kennedy, J. L., & Macciardi, F. (2005). Combined analysis of patients confirms an age-related association of the serotonin 2A receptor gene with tardive dyskinesia and specificity for the non-orofacial subtype. The international journal of neuropsychopharmacology, 8(3), 411-425. https://doi.org/10.1017/S146114570500538
- 127. Li, Y., Song, X., Zhao, X., Zou, L., & Xu, G. (2014). Serum metabolic profiling study of lung cancer using ultra high performance liquid chromatography/quadrupole time-of-flight mass spectrometry. Journal of Chromatography B, 966, 147-153.

9.

- 128. Lieberman, J. A., Stroup, T. S., McEvoy, J. P., Swartz, M. S., Rosenheck, R. A., Perkins, D. O., & Hsiao, J. K. (2005). Effectiveness of antipsychotic drugs in patients with chronic schizophrenia. New England Journal of Medicine, 353(12), 1209-1223.
- 129. Lin, J. H. (2006). Applications and limitations of interspecies scaling and in vitro extrapolation in pharmacokinetics. Drug Metabolism and Disposition, 24(10), 1208-1212.

Llerena, A., Dorado, P., & Peñas-Lledó, E. M. (2014). Pharmacogenetics of CYP2D6 in the metabolism of antipsychotics: clinical implications. Pharmacogenomics, 15(8), 1073-1090.

- 131. Llerena, A., Edman, G., & Cobaleda, J. (1993). Pharmacogenetic aspects of drug metabolism in the elderly. Clinical Pharmacokinetics, 25(2), 99-121.
- 132. Lopez, L. V., & Kane, J. M. (2013). Plasma levels of second-generation antipsychotics and clinical response in acute psychosis: a review of the literature. Schizophrenia Research, 147(2-3), 368-374.
- 133. Luan, J., Yuan, J., Li, X., Jin, S., Yu, L., Liao, M., Zhang, H., Xu, C., He, Q., Wen, B., Zhong, X., Chen, X., Chan, H. L., Sung, J. J., Zhou, B., & Ding, C. (2009). Multiplex detection of 60 hepatitis B virus variants by maldi-tof mass spectrometry. Clinical chemistry, 55(8), 1503–1509.
- 134. Mardal, M., Gracia-Lor, E., Leibnitz, S., Castiglioni, S., & Meyer, M. R. (2016). Toxicokinetics of new psychoactive substances: plasma protein binding, metabolic stability, and human phase I metabolism of the synthetic cannabinoid WIN 55,212-2 studied using in vitro tools and LC-HR-MS/MS. Drug Testing and Analysis, 8(10), 1039-1048.
- 135. Mas, S., Gassó, P., Ritter, M., Malagelada, C., Bernardo, M., & Lafuente, A. (2015). Pharmacogenetic predictor of extrapyramidal symptoms induced by antipsychotics: multilocus interaction in the mTOR pathway. European Neuropsychopharmacology, 25(1), 51-59.
- 136. Matriciani, B., Huppertz, B., Keller, R., & Weiskirchen, R. (2018). False-negative results in the immunoassay analysis of drugs of abuse: Can adulterants be detected by sample check test? Annals of Clinical Biochemistry, 55(3), 348-354.
- 137. Maurer, H. (2017).

 Maurer/Wissenbach/Weber MWW LC-MSn
 Library of Drugs, Poisons, and Their
 Metabolites, 2nd rev. Weinheim, Germany:
 Wiley-VCH.
- 138. Maurer, H. H. (2007). Current role of liquid chromatography–mass spectrometry in clinical and forensic toxicology. Analytical and bioanalytical chemistry, 388, 1315-1325.
- Maurer, H. H. (2010). Analytical toxicology. In Molecular, Clinical and Environmental Toxicology: Volume 2: Clinical Toxicology (pp. 317-338).
- Maurer, H. H. (2018). Mass spectrometry for research and application in therapeutic drug

- monitoring or clinical and forensic toxicology. Therapeutic Drug Monitoring, 40(4), 389-393.
- 141. Maurer, H. H., Pfleger, K., & Weber, A. A. (2007). Mass spectral library of drugs, poisons, pesticides, pollutants and their metabolites, 4th revision. Wiley-VCH, Weinheim.
- 142. Maurer, H., & Meyer, M. R. (2014). Methods for urine drug testing using one-step dilution and direct injection in combination with LC– MS/MS and LC–HRMS. Bioanalysis, 6(17), 2229-2244.
- 143. Maurer, H., & Meyer, M. R. (2016). Highresolution mass spectrometry in toxicology: current status and future perspectives. Archives of toxicology, 90, 2161-2172.
- 144. Mbughuni, M. M., Jannetto, P. J., & Langman, L. J. (2016). Mass spectrometry applications for toxicology. Ejifcc, 27(4), 272.
- 145. McElroy, S. L., Keck Jr, P. E., Strakowski, S. M., & Bourne, M. L. (1997). Clinical and research implications of the diagnosis of dysphoric or mixed mania or hypomania. American Journal of Psychiatry, 154(11), 1633-1644.
- 146. Meijer, D. K. F., & Smit, J. W. (2000). The influence of transporters in pharmacokinetic processes. European Journal of Pharmaceutical Sciences, 12(4), 273-293.
- 147. Menke, A., Klengel, T., & Binder, E. B. (2012). Epigenetics, depression and antidepressant treatment. Current Pharmaceutical Design, 18(36), 5879-5889.
- 148. Meyer, J. M., & Stahl, S. M. (2009). The metabolic syndrome and schizophrenia. Acta Psychiatrica Scandinavica, 119(4), 289-310.
- 149. Meyer, M. R. (2016). New psychoactive substances: an overview on recent publications on their toxicodynamics and toxicokinetics. Archives of toxicology, 90, 2421-2444.
- 150. Meyer, M. R., & Maurer, H. H. (2012). Current applications of high-resolution mass spectrometry in drug metabolism studies. Analytical and bioanalytical chemistry, 403, 1221-1231.
- 151. Meyer, M. R., & Maurer, H. H. (2016). LC coupled to low-and high-resolution mass spectrometry for new psychoactive substance screening in biological matrices—where do we stand today? Analytica chimica acta, 927, 13-20.
- 152. Meyer, M. R., Peters, F. T., & Maurer, H. H. (2010). Automated mass spectral deconvolution and identification system for GC-MS screening for drugs, poisons, and metabolites in urine. Clinical chemistry, 56(4), 575-584.
- 153. Meyer, M. R., Wagmann, L., Schneider-Daum, N., Loretz, B., de Souza Carvalho, C., Lehr, C.

- M., & Maurer, H. H. (2015). P-glycoprotein interactions of novel psychoactive substances stimulation of ATP consumption and transport across Caco-2 monolayers. Biochemical pharmacology, 94(3), 220–226. https://doi.org/10.1016/j.bcp.2015.01.008
- 154. Meyer, R., Gehlhaus, M., Knoth, R., & Volk, B. (2007). Expression and function of cytochrome p450 in brain drug metabolism. Current Drug Metabolism, 8(4), 297-306.
- 155. Michely, J. A., Meyer, M. R., & Maurer, H. H. (2017). Dried urine spots-A novel sampling technique for comprehensive LC-MSn drug screening. Analytica chimica acta, 982, 112-121.
- 156. Michely, J. A., Meyer, M. R., & Maurer, H. H. (2017). Paper spray ionization coupled to high resolution tandem mass spectrometry for comprehensive urine drug testing in comparison to liquid chromatography-coupled techniques after urine precipitation or dried urine spot workup. Analytical chemistry, 89(21), 11779-11786.
- 157. Mills, S., & Lee, D. Y. W. (2017). Pharmacokinetic interactions between conventional medications and herbal medicines. Expert Opinion on Drug Metabolism & Toxicology, 13(3), 317-334.
- 158. Milosheska, D., Grabnar, I., & Vovk, T. (2014). Dried blood spots for monitoring and individualized dosing of antiepileptic drugs. European Journal of Pharmaceutical Sciences, 59, 20-27.
- 159. Mössner, R., Schuhmacher, A., Kühn, K. U., Cvetanovska, G., Rujescu, D., Zill, P., Quednow, B. B., Rietschel, M., Wölwer, W., Gaebel, W., Wagner, M., & Maier, W. (2009). Functional serotonin 1A receptor variant influences treatment response to atypical antipsychotics in schizophrenia. Pharmacogenetics and genomics, 19(1), 91–94. https://doi.org/10.1097/FPC.0b013e328311 a917.
- 160. Müller, M. J., Regenbogen, B., Härtter, S., Eich, F. X., & Hiemke, C. (2007). Therapeutic drug monitoring for optimizing amisulpride therapy in patients with schizophrenia. Journal of Psychiatric Research, 41(8), 673-679.
- 161. Murray, M. (2006). Role of CYP pharmacogenetics and drug—drug interactions in the efficacy and safety of atypical and other antipsychotic agents. Journal of Pharmacy and Pharmacology, 58(7), 871-885.
- 162. Neef, C., Touw, D. J., & Stolk, L. M. (2008). Therapeutic drug monitoring in clinical

research. Pharmaceutical Medicine, 22, 235-244.

- 163. Nielen, M. W., Vissers, J. P., Fuchs, R. E., v. Velde, J. W., & Lommen, A. (2001). Screening for anabolic steroids and related compounds in illegal cocktails by liquid chromatography /time-of-flight mass spectrometry and liquid chromatography /quadrupole time-of-flight tandem mass spectrometry with accurate mass measurement. Rapid Communications in Mass Spectrometry, 15(17), 1577-1585.
- 164. Oda, S., Fukami, T., Yokoi, T., & Nakajima, M. (2015). A comprehensive review of UDPglucuronosyltransferase and esterases for drug development. Drug Metabolism and Pharmacokinetics, 30(1), 30-51.
- 165. O'Dushlaine, C., Ripke, S., Ruderfer, D. M., Hamilton, S. P., Fava, M., Iosifescu, D. V., Kohane, I. S., Churchill, S. E., Castro, V. M., Clements, C. C., Blumenthal, S. R., Murphy, S. N., Smoller, J. W., & Perlis, R. H. (2014). Rare copy number variation in treatment-resistant major depressive disorder. Biological psychiatry, 76(7), 536–541. https://doi.org/10.1016/j.biopsych.2013.10.0 28.
- 166. Pagotto, U., Fanelli, F., & Pasquali, R. (2013). Insights into tandem mass spectrometry for the laboratory endocrinology. Reviews in Endocrine and Metabolic Disorders, 14, 141-141.
- 167. Pan, Y., & Nicolazzo, J. A. (2018). Impact of aging, Alzheimer's disease, and Parkinson's disease on the blood–brain barrier transport of therapeutics. Advanced Drug Delivery Reviews, 135, 62-74.
- 168. Panderi, I., Perez, K., Cao, L., Noble, L., Lombardo, K., Walsh, T. J., & Pantazatos, D. (2017). Assessment of molecular differentiation in FFPE colon adenocarcinoma tissues using PCA analysis of MALDI IMS spectral data. Journal of Applied Bioanalysis, 3(4), 81–97.
- 169. Pasin, D., Cawley, A., Bidny, S., & Fu, S. (2017). Current applications of high-resolution mass spectrometry for the analysis of new psychoactive substances: a critical review. Analytical and bioanalytical chemistry, 409(25), 5821–5836. https://doi.org/10.1007/s00216-017-0441-4.
- 170. Patsalos, P. N., Berry, D. J., Bourgeois, B. F., Cloyd, J. C., Glauser, T. A., Johannessen, S. I., Leppik, I. E., Tomson, T., & Perucca, E. (2008). Antiepileptic drugs--best practice guidelines for therapeutic drug monitoring: a position paper by the subcommission on therapeutic drug monitoring, ILAE

- Commission on Therapeutic Strategies. Epilepsia, 49(7), 1239–1276. https://doi.org/10.1111/j.1528-1167.2008. 01561.x
- 171. Patteet, L., Maudens, K. E., Stove, C. P., Lambert, W. E., Morrens, M., Sabbe, B., & Neels, H. (2015). The use of dried blood spots for quantification of 15 antipsychotics and 7 metabolites with ultra-high performance liquid chromatography tandem mass spectrometry. Drug testing and analysis, 7(6), 502–511. https://doi.org/10.1002/dta.1698.
- 172. Paul, B. D., Martin, K. K., Maguilo Jr, J., & Smith, M. L. (2000). Effects of pyridinium chlorochromate adulterant (urine luck) on testing for drugs of abuse and a method for quantitative detection of chromium (VI) in urine. Journal of analytical toxicology, 24(4), 233-237.
- 173. Paulzen, M., Eap, C.-B., Gründer, G., & Kuzin, M. (2016). Pharmacokinetic interaction between valproic acid, meropenem, and risperidone. Journal of Clinical Psychopharmacology, 36(4), 405-406.
- 174. Peace, M. R., & Tarnai, L. D. (2002). Performance evaluation of three on-site adulterant detection devices for urine specimens. Journal of analytical toxicology, 26(7), 464-470.
- 175. Pelander, A., Ojanperä, I., Laks, S., Rasanen, I., & Vuori, E. (2003). Toxicological screening with formula-based metabolite identification by liquid chromatography/time-of-flight mass spectrometry. Analytical chemistry, 75(21), 5710-5718.
- 176. Pelander, A., Ristimaa, J., & Ojanperä, I. (2010). Vitreous humor as an alternative matrix for comprehensive drug screening in postmortem toxicology by liquid chromatography-time-of-flight mass spectrometry. Journal of Analytical Toxicology, 34(6), 312-318.
- 177. Pelander, A., Ristimaa, J., Rasanen, I., Vuori, E., & Ojanperä, I. (2008). Screening for basic drugs in hair of drug addicts by liquid chromatography/time-of-flight mass spectrometry. Therapeutic drug monitoring, 30(6), 717-724.
- 178. Pérez, V., Gilaberte, I., Fañanás, L., & Portella, M. J. (2000). Predictors of response to antipsychotic treatment in schizophrenia. Journal of Clinical Psychopharmacology, 20(6), 643-649.
- 179. Perry, P. (2001). Therapeutic drug monitoring of antipsychotics. Psychopharmacology Bulletin, 35(3), 19-29.
- 180. Perry, P. J., Sanger, T., & Beasley, C. (1997). Olanzapine plasma concentrations and clinical

- response in acutely ill schizophrenic patients. Journal of Clinical Psychopharmacology, 17(6), 472-477.
- 181. Perry, P. J., Zeilmann, C., & Arndt, S. (1994). Tricyclic antidepressant concentrations in plasma: an estimate of their sensitivity and specificity as a predictor of response. Journal of Clinical Psychopharmacology, 14(4), 230-240
- 182. Peters, F. T. (2007). Stability of analytes in biosamples—an important issue in clinical and forensic toxicology? Analytical and Bioanalytical Chemistry, 388, 1505-1519.
- 183. Peters, F. T. (2011). Recent advances of liquid chromatography—(tandem) mass spectrometry in clinical and forensic toxicology. Clinical Biochemistry, 44(1), 54-65.
- 184. Petrocheilou, M., Samanidou, V., Kovatsi, L., Tsolaki, M., & Papadoyannis, I. (2017). A simple and direct HPLC-DAD method for the simultaneous determination of galantamine, donepezil and rivastigmine in cerebrospinal fluid, blood serum and urine. Journal of Applied Bioanalysis, 3(4), 59–69. https://doi.org/10.17145/jab.17.010
- 185. Phan, H. M., Yoshizuka, K., Murry, D. J., & Perry, P. J. (2012). Drug testing in the workplace. Pharmacotherapy: The Journal of Human Pharmacology and Drug Therapy, 32(7), 649-656.
- 186. Pirmohamed, M., & Park, B. K. (2003). Genetic susceptibility to adverse drug reactions. Trends in Pharmacological Sciences, 24(7), 385-391.
- 187. Pohjola-Sintonen, S., Kivistö, K. T., Vuori, E., Lapatto-Reiniluoto, O., Tiula, E., & Neuvonen, P. J. (2000). Identification of drugs ingested in acute poisoning: correlation of patient history with drug analyses. Therapeutic drug monitoring, 22(6), 749-752.
- 188. Pounder, D. J., & Jones, G. R. (1990). Postmortem drug redistribution—a toxicological nightmare. Forensic science international, 45(3), 253-263.
- 189. Praveen, M., Ullah, I., Buendia, R., Khan, I. A., Sayed, M. G., Kabir, R., Bhat, M. A., Yaseen, M. (2024). Exploring Potentilla nepalensis Phytoconstituents: Integrated Strategies of Network Pharmacology, Molecular Docking, Dynamic Simulations, and MMGBSA Analysis for Cancer Therapeutic Targets Discovery. Pharmaceuticals (Basel). 17(1):134. doi: 10.3390/ph17010134.
- 190. Praveen, M. (2024). Characterizing the West Nile Virus's polyprotein from nucleotide sequence to protein structure Computational tools. J Taibah Univ Med Sci. 19(2):338-350. doi: 10.1016/j.jtumed.2024.01.001.

191. Praveen, M. (2024). Multi-epitope-based vaccine designing against Junín virus glycoprotein: immunoinformatics approach. Futur J Pharm Sci 10, 29.

https://doi.org/10.1186/s43094-024-00602-8

- 192. Praveen, M., Morales-Bayuelo, A. (2023). Drug Designing against VP4, VP7 and NSP4 of Rotavirus Proteins Insilico studies, Mor. J. Chem., 14(6), 729-741.
- 193. Preskorn, S. H. (2014). Therapeutic Drug Monitoring (TDM) in psychiatry (part I): why studies attempting to correlate drug concentration and antidepressant response don't work. Journal of Psychiatric Practice®, 20(2), 133-137.
- 194. Preskorn, S. H., Burke, M. J., Fast, G. A., & Ciardullo, T. (1992). Therapeutic drug monitoring: principles and practice. Psychiatric Clinics of North America, 15(1), 153-175.
- 195. Preskorn, S. H., Flockhart, D., Tamminga, C., Mcdougle, C. J., Guengerich, F. P., Cohen, L., & Guideline, S. F. (2006). Antipsychotic drugdrug interactions: an evidence-based review. Journal of Clinical Psychiatry, 67(Suppl 7), 1-47.
- 196. Proft, F., Kopf, J., Olmes, D., Hempel, S., Schmidt, B., Riederer, P., Deckert, J., Pfuhlmann, B., Reif, A., & Unterecker, S. (2014). SLC6A2 and SLC6A4 variants interact with venlafaxine serum concentrations to influence therapy outcome. Pharmacopsychiatry, 47(7), 245–250. https://doi.org/10.1055/s-0034-1390412.
- 197. Rajkumar, A. P., Christensen, J. H., Nickelsen, M., & Sørensen, T. (2013). Antipsychotic dose conversion between oral and long-acting injectable formulations using a three-compartment model. Journal of Clinical Psychopharmacology, 33(3), 318-321.
- 198. Rao, V. R., Bishop, M., & Coppen, A. (1980). Clinical state, plasma levels of haloperidol and prolactin: a correlation study in chronic schizophrenia. The British Journal of Psychiatry, 137(6), 518-521.
- 199. Rawls, S. M., & Benamar, K. (2011). Dual norepinephrine reuptake inhibition and serotonin receptor blockade for the treatment of mood disorders: therapeutic rationale and efficacy of mirtazapine. Pharmacology & Therapeutics, 129(3), 287-294.
- 200. Reis, M., et al., (2007). Reference concentrations of antidepressants. A compilation of postmortem and therapeutic levels. Journal of Analytical Toxicology, 31(5), 254-264.

201. Remane, D., Meyer, M. R., Wissenbach, D. K., & Maurer, H. H. (2010). Ion suppression and enhancement effects of co-eluting analytes in multi-analyte approaches: systematic investigation using ultra-high-performance liquid chromatography/mass spectrometry with atmospheric-pressure chemical ionization or electrospray ionization. Rapid Communications in Mass Spectrometry, 24(21), 3103-3108.

- 202. Remane, D., Wissenbach, D. K., Meyer, M. R., Maurer, H. H. (2010). Systematic investigation of ion suppression and enhancement effects of fourteen stableisotope-labeled internal standards by their native analogues using atmospheric-pressure chemical ionization and electrospray ionization and the relevance for multi-analyte liquid chromatographic/mass spectrometric procedures. Rapid Communications in Mass Spectrometry: An International Journal Devoted to the Rapid Dissemination of Upto-the-Minute Research in Mass Spectrometry, 24(7), 859-867.
- 203. Richter, L. H., Flockerzi, V., Maurer, H. H., & Meyer, M. R. (2017). Pooled human liver preparations, HepaRG, or HepG2 cell lines for metabolism studies of new psychoactive substances? A study using MDMA, MDBD, butylone, MDPPP, MDPV, MDPB, 5-MAPB, and 5-API as examples. Journal of Pharmaceutical and Biomedical Analysis, 143, 32-42.
- 204. Richter, L. H., Maurer, H. H., & Meyer, M. R. (2017). New psychoactive substances: studies on the metabolism of XLR-11, AB-PINACA, FUB-PB-22, 4-methoxy-α-PVP, 25-I-NBOMe, and meclonazepam using human liver preparations in comparison to primary human hepatocytes, and human urine. Toxicology Letters, 280, 142-150.
- 205. Rochat, B., Kosel, M., Boss, G., Testa, B., Gillet, M., & Baumann, P. (1998). Stereoselective biotransformation of the selective serotonin reuptake inhibitor citalopram and its demethylated metabolites by monoamine oxidases in human liver. Biochemical Pharmacology, 56(1), 15-23.
- 206. Roemmelt, A. T., Steuer, A. E., & Kraemer, T. (2015). Liquid chromatography, in combination with a quadrupole time-of-flight instrument, with sequential window acquisition of all theoretical fragment-ion spectra acquisition: validated quantification of 39 antidepressants in whole blood as part of a simultaneous screening and quantification

procedure. Analytical chemistry, 87(18), 9294-9301.

- 207. Roerig, J. L., Steffen, K. J., & Mitchell, J. E. (2011). Atypical antipsychotic-induced weight gain: insights into mechanisms of action and implications for clinical management. Prostaglandins, Leukotrienes and Essential Fatty Acids, 85(2), 149-156.
- 209. Schoretsanitis, G., Haen, E., Stegmann, B., Hiemke, C., Gründer, G., & Paulzen, M. (2017). Effect of smoking on risperidone pharmacokinetics A multifactorial approach to better predict the influence on drug metabolism. Schizophrenia research, 185, 51–57.

https://doi.org/10.1016/j.schres.2016.12.016

- 210. Schoretsanitis, G., Kane, J. M., & Citrome, L. (2018). Personalized treatment of schizophrenia and the role of pharmacogenetics. The World Journal of Biological Psychiatry, 19(1), 3-11.
- 211. Schoretsanitis, G., Kane, J. M., Citrome, L., Halkin, A., Olfson, M., Newcomer, J. W., ... & Ruschena, R. (2020). Personalized treatment of schizophrenia and the role of pharmacogenetics. The World Journal of Biological Psychiatry, 21(9), 690-703.
- 212. Segman, R. H., Heresco-Levy, U., Finkel, B., Goltser, T., Shalem, R., Schlafman, M., Dorevitch, A., Yakir, A., Greenberg, D., Lerner, A., & Lerer, B. (2001). Association between the serotonin 2A receptor gene and tardive dyskinesia in chronic schizophrenia. Molecular psychiatry, 6(2), 225–229.
- https://doi.org/10.1038/sj.mp.4000842
 213. Segman, R. H., Heresco-Levy, U., Finkel, B., Inbar, R., Neeman, T., Schlafman, M., Dorevitch, A., Yakir, A., Lerner, A., Goltser, T., Shelevoy, A., & Lerer, B. (2000). Association between the serotonin 2C receptor gene and tardive dyskinesia in chronic schizophrenia: additive contribution of 5-HT2Cser and DRD3gly alleles to susceptibility. Psychopharmacology, 152(4), 408–413.

https://doi.org/10.1007/s002130000521.

214. Shah, R. R. (2006). Drug-induced QT interval prolongation—regulatory guidance and perspectives on hERG channel studies. Novartis Foundation Symposium, 277, 251-280.

- 215. Shanks, K. G., Dahn, T., Behonick, G., & Terrell, A. (2012). Analysis of first and second generation legal highs for synthetic cannabinoids and synthetic stimulants by ultraperformance liquid chromatography and time of flight mass spectrometry. Journal of analytical toxicology, 36(6), 360-371.
- 216. Sica, D. A. (2005). Drug interactions with olanzapine. Expert Opinion on Drug Metabolism & Toxicology, 1(2), 323-334.
- 217. Smith, S. W. (2009). Chiral toxicology: it's the same thing only different. Toxicological Sciences, 110(1), 4-30.
- 218. Soldin, S. J., & Soldin, O. P. (2009). Steroid hormone analysis by tandem mass spectrometry. Clinical chemistry, 55(6), 1061-1066.
- 219. Sparshatt, A., Taylor, D., Patel, M. X., & Kapur, S. (2010). A systematic review of aripiprazole—dose, plasma concentration, receptor occupancy, and response: implications for therapeutic drug monitoring. The Journal of Clinical Psychiatry, 71(11), 20680
- 220. Spina, E., & de Leon, J. (2014). Metabolic drug interactions with newer antipsychotics: a comparative review. Basic & Clinical Pharmacology & Toxicology, 114(3), 396-412.
- 221. Stead, A., & Moffat, A. (1983). A collection of therapeutic, toxic and fatal blood drug concentrations in man. Human Toxicology, 2(3), 437-464.
- 222. Stieffenhofer, V., Saglam, H., Schmidtmann, I., Silver, H., Hiemke, C., & Konrad, A. (2011). Clozapine plasma level monitoring for prediction of rehospitalization schizophrenic outpatients. Pharmacopsychiatry, 44(2), 55–59. https://doi.org/10.1055/s-0030-1267178.
- 223. Stove, C. P., Ingels, A.-S. M., De Kesel, P. M., & Lambert, W. E. (2012). Dried blood spots in toxicology: from the cradle to the grave? Critical reviews in toxicology, 42(3), 230-243.
- 224. Takekita, Y., Fabbri, C., Kato, M., Koshikawa, Y., Tajika, A., Kinoshita, T., & Serretti, A. (2016). HTR1A Polymorphisms and Clinical Efficacy of Antipsychotic Drug Treatment in Schizophrenia: A Meta-Analysis. The international journal of neuropsychopharmacology, 19(5), pyv125. https://doi.org/10.1093/ijnp/pyv125
- 225. Taurines, R., Burger, R., Wewetzer, C., Pfuhlmann, B., Mehler-Wex, C., Gerlach, M., & Egberts, K. (2013). The relation between

- dosage, serum concentrations, and clinical outcome in children and adolescents treated with sertraline: a naturalistic study. Therapeutic drug monitoring, 35(1), 84–91.
- https://doi.org/10.1097/FTD.0b013e31827a 1aad
- 226. Tenore, P. L. (2010). Advanced urine toxicology testing. Journal of addictive diseases, 29(4), 436-448.
- 227. Thorn, C. F., & Klein, T. E. (2011). Altman RB Pharmacogenomics and bioinformatics: PharmGKB. Pharmacogenomics, 12(8), 1053-1056.
- 228. Thummel, K. E., & Wilkinson, G. R. (1998). In vitro and in vivo drug interactions involving human CYP3A. Annual Review of Pharmacology and Toxicology, 38, 389-430.
- 229. Uebel, R. A., & Wium, C. A. (2002). Toxicological screening for drugs of abuse in samples adulterated with household chemicals. South African Medical Journal-Cape Town-Medical Association Of South Africa-, 92(7; PART 1), 547-548.
- 230. Ulrich, S., & Läuter, J. (2002). Comprehensive survey of the relationship between serum concentration and therapeutic effect of amitriptyline in depression. Clinical Pharmacokinetics, 41, 853-876.
- 231. Ulrich, S., Wurthmann, C., Brosz, M., & Meyer, F. P. (1998). The relationship between serum concentration and therapeutic effect of haloperidol in patients with acute schizophrenia. Clinical Pharmacokinetics, 34, 227-263.
- 232. United Nations Office on Drugs and Crime. (2018). Early warning advisory on new psychoactive substances.
- 233. Unterecker, S., et al., (2015). Increase of heart rate and QTc by amitriptyline, but not by venlafaxine, is correlated to serum concentration. Journal of Clinical Psychopharmacology, 35 (4), 460-463.
- 234. Urban, J. D., Benvenga, M. J., & Dillon, M. (2010). Drug–drug interactions involving p-glycoprotein substrates: therapeutic implications. Current Drug Metabolism, 11(8), 685-693.
- 235. Valtier, S., & Cody, J. T. (2002). A Procedure for the Detection of StealthTM Adulterant in Urine Samples. Clinical Laboratory Science, 15(2), 111-115.
- 236. VanderZwaag, C., McGee, M., McEvoy, J. P., Freudenreich, O., Wilson, W. H., & Cooper, T. B. (1996). Response of patients with treatment-refractory schizophrenia to clozapine within three serum level ranges. The

American Journal of Psychiatry, 153 (12), 1579-1584.

- 237. Verplaetse, R., & Henion, J. (2016). Quantitative determination of opioids in whole blood using fully automated dried blood spot desorption coupled to on-line SPE-LC-MS/MS. Drug testing and analysis, 8(1), 30-38.
- 238. Verstraete, A. G. (2004). Detection times of drugs of abuse in blood, urine, and oral fluid. Therapeutic drug monitoring, 26(2), 200-205.
- 239. Virus, E., Sobolevsky, T., & Rodchenkov, G. (2008). Introduction of HPLC/orbitrap mass spectrometry as screening method for doping control. Journal of mass spectrometry, 43(7), 949-957.
- 240. Vogeser, M., & Parhofer, K. (2007). Liquid chromatography tandem-mass spectrometry (LC-MS/MS)-technique and applications in endocrinology. Experimental and clinical endocrinology & diabetes, 115(09), 559-570.
- 241. von Mach, M.-A., Weber, C., Meyer, M. R., Weilemann, L. S., Maurer, H. H., & Peters, F. T. (2007). Comparison of urinary on-site immunoassay screening and gas chromatography-mass spectrometry results of 111 patients with suspected poisoning presenting at an emergency department. Therapeutic drug monitoring, 29(1), 27-39.
- 242. Vuori, E., & Ojanperä, I. (2009). Forensic applications of toxicology. In Wiley encyclopedia of forensic science-Wiley-Blackwel, 2009 (pp. 2503-2509).
- 243. Waldschmitt, C., Vogel, F., Pfuhlmann, B., & Hiemke, C. (2009). Duloxetine serum concentrations and clinical effects. Data from a therapeutic drug monitoring (TDM) survey. Pharmacopsychiatry, 42 (05), 189-193.
- 244. Whiteaker, J. R. (2010). The increasing role of mass spectrometry in quantitative clinical proteomics. Oxford University Press, 56, 1373-1374.
- 245. Williams, J. A., Hyland, R., Jones, B. C., Smith, D. A., Hurst, S., Goosen, T. C., & Ball, S. E. (2004). Drug–drug interactions for UDP-glucuronosyltransferase substrates: a pharmacokinetic explanation for typically observed low exposure (AUCI/AUC) ratios. Drug Metabolism and Disposition, 32(11), 1201-1208.
- 246. Williamson, B., & Aaronson, S. (2016). Medication adherence in schizophrenia: the role of cognitive functioning and insight. Journal of Clinical Psychopharmacology, 36(4), 380-382.
- 247. Wissenbach, D. K., Meyer, M. R., Remane, D., Philipp, A. A., Weber, A. A., & Maurer, H. H. (2011). Drugs of abuse screening in urine as part of a metabolite-based LC-MSn screening

concept. Analytical and bioanalytical chemistry, 400(10), 3481–3489.

- https://doi.org/10.1007/s00216-011-5032-1.
- 248. Wissenbach, D. K., Meyer, M. R., Remane, D., Weber, A. A., & Maurer, H. H. (2011). Development of the first metabolite-based LC-MS(n) urine drug screening procedure-exemplified for antidepressants. Analytical and bioanalytical chemistry, 400(1), 79–88. https://doi.org/10.1007/s00216-010-4398-9.
- 249. Wissenbach, D. K., Meyer, M. R., Weber, A. A., Remane, D., Ewald, A. H., Peters, F. T., & Maurer, H. H. (2012). Towards a universal LC-MS screening procedure can an LIT LC-MS(n) screening approach and reference library be used on a quadrupole-LIT hybrid instrument?. Journal of mass spectrometry: JMS, 47(1), 66–71. https://doi.org/10.1002/jms.2027.
- 250. Wohkittel, C., Gerlach, M., Taurines, R., Wewetzer, C., Unterecker, S., Burger, R., Schreck, D., Mehler-Wex, C., Romanos, M., & Egberts, K. (2016). Relationship between clozapine dose, serum concentration, and clinical outcome in children and adolescents in clinical practice. Journal of neural transmission (Vienna, Austria: 1996), 123(8), 1021–1031. https://doi.org/10.1007/s00702-016-1573-y
- 251. Wu, A. H., Bristol, B., Sexton, K., Cassella-McLane, G., Holtman, V., & Hill, D. W. (1999). Adulteration of urine by "Urine Luck". Clinical Chemistry, 45(7), 1051-1057.
- 252. Wu, M.-K., Chung, W., Wu, C.-K., & Tseng, P.-T. (2015). The severe complication of Stevens–Johnson syndrome induced by long-term clozapine treatment in a male schizophrenia patient: a case report. Neuropsychiatric Disease and Treatment, 11, 1039-1041.
- 253. Xu, X. (2016). In vivo characterization of therapeutic monoclonal antibodies. Journal of Applied Bioanalysis, 2(1), 10–15.
- 254. Wyman, J. F. (2012). Principles and procedures in forensic toxicology. Clinics in laboratory medicine, 32(3), 493-507.
- 255. Yáñez, J. A., Remsberg, C. M., Sayre, C. L., Forrest, M. L., & Davies, N. M. (2011). Flip-flop pharmacokinetics—delivering a reversal of disposition: challenges and opportunities during drug development. Therapeutic Delivery, 2(5), 643-672.
- 256. Yasui-Furukori, N., et al., (2010). Clinical response to risperidone in relation to plasma drug concentrations in acutely exacerbated schizophrenic patients. Journal of Psychopharmacology, 24 (7), 987-994.
- 257. Yin, O. Q., et al., (2006). Phenotype-genotype relationship and clinical effects of citalogram

in Chinese patients. Journal of clinical psychopharmacology, 26(4), 367-372.

- 258. Yuan, C., Chen, D., & Wang, S. (2015). Drug confirmation by mass spectrometry: Identification criteria and complicating factors. Clinica Chimica Acta, 438, 119-125.
- 259. Zanger, U. M., & Schwab, M. (2013). Cytochrome P450 enzymes in drug metabolism: regulation of gene expression, enzyme activities, and impact of genetic variation. Pharmacology & Therapeutics, 138(1), 103-141.
- 260. Zhou, S.-F. (2009). Polymorphism of human cytochrome P450 2D6 and its clinical significance: part II. Clinical Pharmacokinetics, 48, 761-804.
- 261. Zhou, S.-F., Liu, J.-P., & Chowbay, B. (2009). Polymorphism of human cytochrome P450 enzymes and its clinical impact. Drug Metabolism Reviews, 41(2), 89-295.